ISSN: 0975 -8542



Journal of Global Pharma Technology

Available Online at: www.jgpt.co.in

RESEARCH ARTICLE

Evaluation of Analgesic Activity of Extracts of *Jasminum* sessiliflorum Vahl

Ruby Philip^{1*}, Kathiresan Krishnasamy², Elessy Abraham³

- ^{1.} Nazareth College of Pharmacy, Othera, Thiruvalla, Kerala.
- ² Department of Pharmacy, Annamalai University, Annamalai Nagar, Chidambaram.
- 3. Nazareth College of Pharmacy, Othera, Thiruvalla, Kerala.

*Corresponding Author: Ruby Philip

Abstract

The present study was designed to investigate analgesic activity of ethyl acetate and ethanolic extracts of *Jasminum sessiliflorum* Vahl. The analgesic activity was assessed by using acetic acid induced writhing method and tail immersion tests in rats. The ethyl acetate and ethanolic extracts decreased the acetic acid induced abdominal contractions and also significantly increased the reaction time of tail immersion test [1]. These results showed that the plant extracts had both central and peripheral analgesic action. The results suggest that the extracts of *J.sessiliflorum* possesses potent analgesic properties, which support its use in traditional medicine and suggesting that the plant should be further investigated for its pharmacological active natural products [2].

Keywords: Analgesic activity, J. sessiliflorum, Acetic acid induced writhing, Tail immersion test.

Introduction

Algesia or pain is an ill-defined, unpleasant sensation, usually evoked by an external or internal noxious stimulus. An analgesic selectively relieves pain by acting in the CNS or on peripheral pain mechanisms, without significantly altering consciousness. So analgesic activity means capacity of a substance to neutralize the pain sensation [3].

Pain is an important signal and primarily protective in nature which causes discomfort. Analgesics relieve pain as a symptom, without affecting its cause [4]. Analgesics are classes of drugs used to relieve pain.

The pain relief induced by analgesics occurs either by blocking pain signals going to the brain or by interfering with the brain's interpretation of the signals, without producing anesthesia or loss of consciousness. Analgesic activity of drug or any test sample at different steps of pharmacological investigation can be assessed by various methods like acetic acid induced writhing method and tail immersion tests.

The acetic acid induced writhing method is an analgesic behavioral observation assessment method that demonstrates a noxious stimulation in mice. In tail immersion tests, the flick of the tail when immersed in boiling water is recorded.

Jasminum sessiliflorum Vahl is a species of jasmine native to India, Sri Lanka and the Andaman Islands. It is a climbing shrub with a smooth stem and minutely pubescent branchlets.

Medicinal plants have been valued in developing countries of the world for primary health care due to better cultural acceptability, better compatibility with human body and lesser number of side effects [5].

Materials and Methods

Collection of Plant Materials

The plant J. sessiliflorum Vahl (Family:

Oleaceae) were collected from Tirunelveli district, Tamilnadu, India. The plant was identified and authenticated by Mr. Chelladurai.

Preparation of Extracts

About 1 kg of air-dried plant *Jasminum* sessliflorum, was extracted in soxhlet assembly successively with petroleum ether, chloroform, ethyl acetate, and ethanol (order of increasing polarity). Hot percolation method was employed to obtain the aqueous extract. Each time before extracting with the next solvent, the powdered material was dried at room temperature.

Each extract was concentrated by using rotary vacuum evaporator. The extract obtained with each solvent was weighed and the percentage yield was calculated in terms of dried weight of the plant material. The colour and consistency of the extract were also noted. All the solvents used for this entire work were of analytical reagent grade (Merck, Mumbai).

Animals

28 adult male albino rats (180-220 g) were housed in animal house under standard laboratory conditions (temperature $23 \pm 2\,^{\circ}\text{C}$) with 12 h dark and 12 h light cycle. The animals had free access to standard dry pellet diet and water ad libitum. Permission for conduct of these experiments was obtained from, Institutional Animal Ethics Committee.

Antinociceptive Activity

Acetic Acid Induced Writhing Test

The abdominal writhing response to the acetic acid administration involves contractions of the hind limbs. The number of abdominal writhing was recorded for a period of 10 min, starting 5 min after the administration [7]. For the writhing tests, rats received acetic acid injection 30 min after receiving their respective treatment. Group I was pretreated with distilled water.

Group II received diclofenac sodium (10mg/kg b.w, i.p). Group III, IV, V, VI were treated with ethyl acetate and ethanolic extracts (200 mg/kg b.w o.d; 400 mg/kg b.w, orally) respectively [8]. The percentage analgesic activity (PAA) was calculated by using the following formula:

$$PAA = \frac{(C - CD)}{CD} X 100$$

C = Mean of contractions count in animals treated with different doses of ethyl acetate and ethanolic extract of *J.sessiliflorum* and Diclofenac sodium CD = Mean of contractions count in animals served as negative control

Tail Immersion Tests in Rats

The tail immersion test basically involves the measure of the response latency of rats to a nociceptive stimulus. This involves introducing 3 cm of the rat's tail in hot water at a temperature of $55 \pm 0.5^{\circ}$ C. The reaction time taken by the animal to withdraw its tail was recorded (using a chronometer). The time reaction is taken at 30, 60, 90 and 120 administration after of different preparation [9].

Statistical Analysis

The results are reported as mean \pm S.E.M. The statistical analyses were performed using one way analysis of variance (ANOVA). Group differences were calculated by post hoc analysis using Tukey's test. For all tests, differences with values of P<0.05 were considered significant

Results and Discussions

Antinociceptive Activity

Acetic Acid Induced Writhing Test

The results presented in Table 1, shows that the ethyl acetate and ethanolic extracts (200 and 400 mg/kg b.w) of *Jasminum sessiliflorum Vahl* inhibited significantly (p< 0.001) the acetic acid induced abdominal constrictions.

The protective effect of the extracts reached a maximum inhibition of 61.96% for ethanolic extract and 56.24% for ethyl acetate extract at the dose of 400 mg/kg b.w. Diclofenac sodium (standard) was more potent than the antinociceptive dose of the extract, with percentage protection of 81.75%.

Table 1: Effects of ethyl acetate and ethanolic extract of J.sessiliflorum on acetic acid-

induced writhing response

Groups	Treatment	Number of writhing	% reduction in
		movements	reaction time
Group I	Distilled water	32.00 ± 2.52	-
Group II	Diclofenac sodium 10mg/kg	5.84 ± 0.90	81.75%
Group III	200mg/kg ethyl acetate extract	16.24 ± 1.68	47.75%
	of J.sessiliflorum		
Group IV	400mg/kg ethyl acetate extract	14.44 ± 1.56	56.24%
	of J.sessiliflorum		
Group V	200mg/kg ethanolic extract of	13.04 ± 1.38	60.48%
	J.sessiliflorum		
Group VI	400mg/kg ethanolic extract of	12.55 ± 1.16	61.96%
	J.sessiliflorum		

Values are expressed as mean \pm SEM

Tail Immersion Test in Rats

The effects of the ethyl acetate and ethanol extracts (200 and 400 mg/kg b.w) of *Jasminum sessiliflorum* in the tail immersion test in rats is presented in Table 2. A significant (p< 0.05) increase in the reaction

time was observed in the tail immersion test in rats after 30, 60, 90 and 120 min when compared with the control. Morphine (standard) at dose of 10 mg/kg b.w exhibited more potent activity at 30, 60, 90 and 120 min than the extracts.

Table 2: Effect of ethanolic and ethylacetate extract of Jasminum sessiliflorum on tail withdrawal reflex

Groups	Drug (dose),	Reaction time (s)				
	route	30 min	60 min	90 min	120 min	
Group I	Distilled water	1.5 ± 0.4	1.6 ± 0.4	1.8 ± 0.4	2.4± 0.4	
Control Group II	10mg/kg (i.p)	5.9 ± 0.2	8.2 ± 0.6	8.5 ± 0.4	8.8 ± 0.6	
Standard	Morphine			3.3		
Group III Ethyl acetate extract	200mg/kg through orally	5.1 ± 0.1	5.4 ± 0.2	5.7 ± 0.4	6.0 ± 0.2	
Group IV Ethyl acetate extract	400mg/kg through orally	5.6 ± 0.2	5.9 ± 0.6	6.2 ± 0.4	6.5 ± 0.8	
Group IV Ethanolic extract	200mg/kg through orally	5.5 ± 0.4	5.9 ± 0.6	6.3 ± 0.4	6.6 ± 0.7	
Group V Ethanolic extract	400mg/kg through orally	6.4 ± 0.6	6.8 ± 0.7	7.2 ± 0.6	7.6 ± 0.8	

Conclusion

The peripheral analgesic effect of the extracts was tested by using acetic acid-induced writhing test. This assay is widely accepted as a model for visceral pain [10, 11]. The involvement of central mechanisms was studied by using the tail-immersion tests, known to activate spinal nociceptive pathway [11].

Ethyl acetate and ethanolic extract of *J. sessiliflorum* demonstrated a dose-dependent, significant antinociceptive activity in animal models of pain. Acetic acid believed to increase the PGE2 and PGF2α in peritoneal fluid (6). The analgesic activity shown in models of pain is indicative that ethyl acetate and ethanolic extracts of *J. sessiliflorum*

might possess centrally and peripherally mediated antinociceptive properties. Chemical components of ethyl acetate and ethanolic extract of *J. Sessiliflorum* such as flavonoids, saponins or phenolic compounds may be responsible for the antinociceptive activities of this extracts.

It can be concluded that the extracts possesses anti-nociceptive properties which are probably mediated via inhibition of prostaglandin synthesis as well as central inhibitory mechanisms which may be of potential benefit for the management of pain and inflammatory disorders. Further investigations are needed to throw light on the potential chemical entity responsible for producing the effects [12, 20].

References

- 1. Domaj MI, Glassco W, Aceto MD and Martin BR (1999) Antinociceptive and pharmacological effects of metanicotina, a selective nicotine agonist. J. Pharmacol. Exp. Ther., 291: 390-398.
- 2. Farshchi A, Ghiasi G, Malek Khatabi P, Farzaei Hossein NA (2009) Antinociceptive Effect of Promethazine in Mice. Iran. J. Basic Med. Sci., 12: 140-145.
- 3. Abdollahi M, Karimpour H, Monsef-Esfehani HR (2003) Antinociceptive effects of Teucrium polium L. total extract and essential oil in mouse writhing test. Pharmacol. Res., 48: 31-35.
- 4. Aoki M, Tsuji M, Takeda H, Harada Y, Nohara J, Matsumiya T, Chiba H (2006) Antidepressants enhance the antinociceptive effects of carbamazepine in the acetic acid-induced writhing test in mice. Europ. J. Pharmacol., 550: 78-83.
- 5. Golshani S, Karamkhani F, Monsef-Esfehani HR, Abdollahi M (2004) Antinociceptive effects of the essential oil of Dracocephalum kotschyi in the mouse writhing test. J. Pharm. Pharm. Sci., 7: 76-79.
- 6. Krasteva I, Momekov G, Zdraveva P, Konstantinov S, Nikolov S (2008) Antiproliferative effects of a flavonoid and saponins from Astragalus hamosus against human tumor cell lines. Pharmacognosy Magazine, 4: 269.
- 7. Bektas N, Arslan R (2010) Antinociceptive effects of methanol extract of Capparisovate in mice. Pharm Biol., 48: 1185-1190.
- 8. Koster R, Anderson M, De Beer J (1959) Acetic acid for analgesic screening. Fed. Proc., 18: 412-417.
- Aydin S, Demir T, Ozturk Y, Husnu K, Baser C (1999) Analgesic activity of Nepeta italica L. Phytother Res., 13: 20-23.
- 10. Khanna P, Sharma OP, Seghal M, Bhargava C, et al (1980) Antimicrobial principles from in vivo tissue culture of some species. Ind. J. of Pharma. Sci., 42: 113-117.

- 11. D'Amour FE, Smith DL (1941) A method for determining loss of pain sensation. J. Pharmacol. & Exp. Therap., 72: 74-79
- 12. Paulino N, Dankas AP, Bankova V, Longhi DT, Scremin A de Castro SL, Calixto JB (2003) Bulgarian propolis induces analgesic and anti-inflammatory effects in mice and inhibits airway smooth muscle. J. Pharmacol. Sci., 93: 307-313.
- 13. Tjolsen A, Berge OG, Hunskaar S, Rosland JH, Hole K (1992) The formalin test: an evaluation of the method. Pain, 51: 5-17.
- 14. Paul S, Saha D (2012) Analgesic activity of methanol extract of Plumbago indica (L.) by acetic acid induced writhing method. Asian J. Pharm. Tech., 2(2):74-76.
- 15. Kulkarni SK (1999) Handbook of Experimental Pharmacology. New Delhi: Vallabh Prakashan, Revised Edition 3rd, 123-25.
- 16. Tripathi KD (2001) Essentials of medical Pharmacology. Medical Publishers (P) Ltd, New Delhi: 6th Edition, 420.
- 17. Jamal AK, Yaacob WA, Din LB (2008) A Chemical study on Phyllanthus reticulatus. Journal of Physical Science, 19(2): 45-50.
- 18. Harborne JB (1998) Phytochemical methods: a guide to modern techniques of plant analysis, 2nd edition, London: Chapmand and Hall, 54-84.
- 19. Guide for care and use of laboratory animals (1959) National Academy of Sciences Committee for the Update of the Guide for the Care and Use of Laboratory Animals, the National Academies P. 22. Koster R, Anderson M, Rehan HMS. Acetic acid analgesic screening. Federation proceeding 18: 418.
- 20. Taber RI, Greenhouse DD, Rendel JK, Irwin S (1969) Agonist and antagonist interaction of opoids on acetic acid induced abdominal stretching in mice. J. Pharmacol., 169: 29-37. Ress, 8e edition. 2011.