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**RESEARCH ARTICLE** 

Determination of Mefenamic Acid using 8-hydroxy quinoline as a Precipitating Agent and Low Pressure Mercury Lamp (184.9 & 253.7 nm) as a Source of Irradiation using of ISNAG Continue Flow Fluorimeter

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#### Abstract

A new, simple, sensitive and fast method for the determination of Mefenamic acid in pure form and drugs (tablets) by continuous flow feed analysis via measurement of diverged beam of light by ISNAG\_fluorimeter (homemade instrument). The method based on the reaction of the Mefenamic acid with 8-HQ to form yellow precipitate. Optimum parameters have been studied to increase the sensitivity for the developed method. The linear dynamic range for the instrument response versus Mefenamic acid concentration was 0.005-7 mmol/L while the L.O.D was 0.216 µg/sample from the step wise dilution for the minimum concentration of lowest concentration in the linear dynamic range of the calibration graph. The correlation coefficient (r) was 0.9956 while percentage linearity (R²%) was 99.12%. RSD % for the repeatability (n=8) was lower than 0.2% for the determination of Mefenamic acid, with concentration of 2, 5 mmol/L respectively. The developed method was applied successfully for the determination of Mefenamic acid in pharmaceutical tablets. A comparison was made between the newly developed method with the classical method (UV-Vis spectrophotometry at wavelength 285nm, and Turbidemtric method) of analysis using the standard addition method via the use of paired t-test. It shows that there was no significant difference between the quoted values of each individual company with calculated t-value at 95% confidence interval from developed method.

**Keywords:** Mefenamic acid, Flow injection analysis, Homemade instrument.

#### Introduction

(MFNC) Mefenamic acid [(2,3-dimethylphenyl) aminol benzoic acid and is used as an analgesic and anti-inflammatory agent (Fig. 1) [1, 4]. It was discovered and brought to market by Parke-Davis in the 1960s. It became generic in the 1980s and is available worldwide under many brand names [5-7]. Scientists led by Claude Winder from Parke-Davis invented Mefenamic acid in 1961, along with fellow members of the of anthracitic acid derivatives, flufenamic acid in 1963

and meclofenamate sodium in 1964. U.S. Patent 3,138,636 on the drug was issued in 1964[8-11]. It was approved in the UK in 1963 as "Ponstan", in West Germany in 1964 as "Ponalar", and in France as "Ponstyl" and the US in 1967 as "Ponstel"[12]. Ponstan (Mefenamic acid) is a member of the fenamate group of nonsteroidal anti-inflammatory drugs (NSAIDs). Each bluebanded, ivory capsule contains 250 mg of Mefenamic acid for oral administration [13-16].

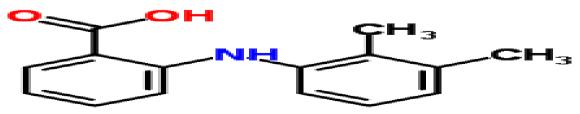


Fig. 1: Chemical structure of Mefenamic acid

Numerous analytical methods have been developed for the determination Mefenamic acid in pure from, dosage forms and biological fluids. Mefenamic acid is official in both United States Pharmacopoeia and British Pharmacopoeia [3, 5, 9]. The USP recommends a high performance liquid chromatographic (HPLC) method for the determination of Mefenamic acid both in raw materials and dosage forms while the BP recommends an acid-base titration method for the analysis of Mefenamic acid in raw materials and its dosage forms [2,8].

Mefenamic acid is a white to greyish-white, odorless, microcrystalline powder with a melting point of 230°-231°C and water solubility of 0.004% at pH 7.1. The chemical name is N-2, 3-xylylanthranilic acid. The molecular weight is 241.29. Its molecular formula is C<sub>15</sub>H<sub>15</sub>NO<sub>2</sub> and the structural formula of Mefenamic acid is: Each capsule also contains lactose, NF. The capsule shell and/or band contains citric acid, USP; D&C yellow No. 10; FD&C blue No. 1; FD&C red No. 3; FD&C yellow No. 6; gelatin, NF; glycerol monolete; silicon dioxide, NF; sodium benzoate, NF; sodium lauryl sulfate, NF; titanium dioxide, USP [17,19].

Several spectrophotometric methods have been reported forthe estimation Mefenamic acid using different reagents such as diazotized 4-amino-3,5- dinitrobenzoicacid [20], methyl-2-benzo-thiazolinone hydrazon hydrochloride after oxidation with Ce+4 or Fe<sup>+3</sup> [21], sodium nitroprusside in the presence of hydroxyl ammonium chloride [22], p-dimethyl amino benzaldehyde [23], pdimethyl amino cinnamaldehyde [24], triton X-114 [25], Fe<sup>+3</sup> to form coloured complex [26]. Many methods have been used in simultaneous determination of Mefenamic acid in the presence of another drug such as paracetamol [27], ethamsylate [28].

However some of these procedures suffer from one or another disadvantage such as: the product may be extracted to organic solvent [20], or require nonaqueous medium [24]and other required control temperature [23, 25]. In this work using continue flow injection scattering method, the diverged beam of light is measured at 0-90° angle will be detected by homemade ISNAGfluorimeter via low-pressure mercury lamp as a source and using 2 sides [4 x 2.5cm] solar cells.

#### **Experimental**

#### Reagent and Chemical

All chemicals were use of analytical-reagent and distilled water was use to prepare all the solutions. A standard solution 5mmol/L of Mefenamic acid molecular formula C<sub>15</sub>H<sub>15</sub>NO<sub>2</sub>, molecular weight 241.29g/mole was prepared by dissolving 0.120645g in 100 ml of NaOH. A stock solution 0.2Mol/L of 8hydroxyquinoline molecular formula (C<sub>9</sub>H<sub>7</sub>NO) molar mass 145.16g/mole and Merck-USA was prepared by dissolving 9.0725g in 250 ml of 0.1Mol/L acetic acid.

#### **Sample Preparation**

Twenty tablets were weight then crushed and mixed. tablet containing 500mg of Mefenamic acid were weighted 0.17723g, 0.17610g, 0.20288, 0.14445g (equivalent to 0.120645g of active ingredient, 5mmol/L) for Ponstan-forte (German)- 500mg, Ponstan-forte (Egypt)-500mg, Merfile-500(India)-500mg, and ponstidin-500(Iraq)-500mg respectively. Each one from the four kinds of sample dissolved in NaOH. The solution was filtered to get rid of undissolved materials, the residue was washed with NaOH and completed the volume to 100ml with the same solvent (NaOH).

#### Apparatus

Two line manifold design to figure out the use of precipitating agent and Mefenamic acid. Figure 2 shows the details of the manifold feed used. ISNAG- fluorimeter was used as a fluorimetric instrument for measurement of total diverged light, which is composed of 100mm distance flow cell at 2mm path length with 2sides [4 x 2.5cm] solar cells. Four on each side i.e.; measure fluorescence at +90° and -90°, which is a novel fluorimetric measurement.

Low-pressure mercury lamp as the excitation source characterized by two strong bands: 184.9nm and 253.7nm, which cannot be detected by ISNAG- fluorimeter unless the fluorescence excitation emission is beyond nm (characteristic of solar cell). Peristaltic pump two channels variable speed (Ismatec, Switzerland). Valve 6-port medium pressure injection valve (IDEX corporation , USA ) with sample loop (1mm i.d. Teflon ,variable length ). The output signals were potentiometric recorder recorded by (Siemens, Germany) (1- 5 Volt, 1000-5000 mV).

Peak height was measured for each signal. Absorbance readings by uv-spectrometer, UV-spectrophotometer (UV-1800 Shimadzu) (Japan). The scattered beam of light for precipitate measured by turbidometry via Turbidity-meter, HANNA- Hungary.

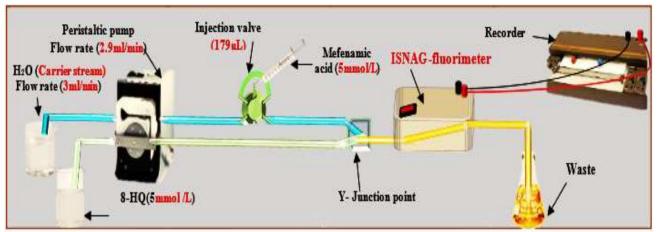


Figure 2: Schematic diagram of flow injection instrument analysis system for determination of Mefenamic acid Fig. 2: Two line flow feed unit

#### Methodology

Using two line system (fig. 2) for the determination of Mefenamic acid with expected parameters of Mefenamic acid (5mmol/L)- 8-HQ (5mmol/L) system, 179µL sample size which will be injected on a

carrier stream line (distilled water) at 3 ml/min while the second line 8-HQ at 2.9ml/min. The precipitate is expected to be probably ion paired compound as the suggested reaction below shows that in scheme 1.

Scheme 1: Proposed mechanism for the reaction between Mefenamic acid and 8-HQ

# Result and Discussion Study of the Optimum Parameters

The flow injection manifold system as shown in Fig.2 was investigated in the relation of chemical and physical variables, in order to obtain optimum conditions for the reaction of 8-HQ with Mefenamic acid and formation of yellow precipitate. They were optimized by making all variables constant and varying one at a time i.e. fixed variable optimization.

#### Variation of Chemical Parameters

Through this section, all variables that contribute in the reaction for determination of Mefenamic acid will be studying for given a best diverged light response measured at 0-90° using four solar cells at each side.

#### 8-hydroxy Quinolone Concentration

At a selected concentration of Mefenamic acid (5mmol/L) was chosen for a set of variable 8-HQ concentration (3-20) mmol/L. No electronic filter was introduced through the measurement of variable concentration. Fig. 3-A shows a kind of responses emission versus time profile; the results of which are tabulated in table no. 1.It was noticed that 5mmol/L of 8-HQ concentration is the most favored concentration to use above this concentration (fig. 3-B); a decrease in diverged light occur; as lower responses were

obtained which might be attributed to the formation of tiny minute particulates or a decrease in the interfacial distances i.e.; prevention of the passage of dispersed and diverged light to the detector.

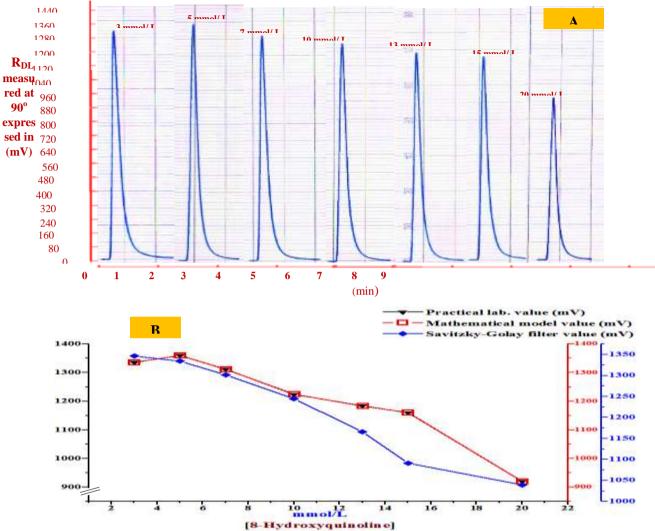


Figure 3:

- Response -time profile of Mefenamic acid with variable concentration of 8-HQ solution (clear un obstructed peak) with no deformation at variable level of 8-HQe solution at (3-20) mmol/L.
- Plot of averaged peak height responses vs. 8-HQ concentration.

Table 1: Effect of 8-HQ concentration on response function expressed as an average peak heights  $\bar{y}i$  (n=3) and tabulation of all available data obtained practically, calculated as obtained by best fit mathematical model, and smoothed digital filtering using Savitzky-Golay data treatment

Independent Dependent variable Average (n=3) diverged light response measured at 0-90° expressed in mV variable [8-HQ] Practical lab. value Mathematical model Savitzky-Golay mmol/L filter Reliability(two tailed)  $\hat{\mathbf{y}}_{i}$ Average peak RSD% ŷi (S-G) height (ȳi)  $\bar{y}_{i}(mV) \pm t_{0.025,2} \sigma_{n-1} / \sqrt{n}$ 3 1336 0.07  $1336 \pm 2.36$ 1335.992 1346.388  $1360 \pm 2.44$ 1360.108 1334.1755 1360 0.07 7 1312 0.08  $1312 \pm 2.71$ 1311.661 1301.644 10 1224 0.09  $1224 \pm 2.78$ 1224.658 1244.455 13 1184 0.07  $1184 \pm 3.28$ 1183.132 1165.632**15** 1160 0.11 $1160 \pm 3.03$ 1160.4851091.126919.964 920 0.16 $920 \pm 3.60$ 1039.338

$$t_{0.025,2} = 4.303$$
,  $\tilde{Y}j = \sum_{i=-\frac{m-1}{2}}^{\frac{m-1}{2}} (Ci \tilde{y}j + i)$ , m= convolution coefficient,  $\frac{m+1}{2} \le j \le n - \frac{m-1}{2}$ 

#### **Carrier Stream Effect**

#### **Acidity Effect**

Fixing Mefenamic acid at 5mmol/L.A series of different acids (H<sub>2</sub>SO<sub>4</sub>, HCl, HNO<sub>3</sub>, and

CH<sub>3</sub>COOH (50mmol/L)) were prepared. Also distilled water was used as a reference. It was noticed a decrease in response profile which might be attributed to the dissociation and dispersed precipitated particles and

minimum large precipitated particles that causes the diverged light beam due to increased reflecting surface area. Therefore, distilled water was chosen as the most satisfactory medium. The overall results are tabulated in Table 2.

 $Table~2:~Effect~of~acidity~medium~as~a~carrier~stream~on~height~of~responses~expressed~as~an~average~peak~heights~\bar{y}_i$ 

(n=3) using 79µL from MFE (5mmol/L) and 8-HQ (5mmol/L)

Independent type of acid as a carrier stream [acid] 50mmol/L	Dependent variable (ỹ <sub>i</sub> ) Average (n=3) diverged light response measured at 0- 90° expressed in mV	RSD%	Reliability of average response(two tailed) at 95% confidence level \$\bar{v}_i\$ (mV)\pm t_{0.025,2} \(\sigma_{n-1}/\sqrt{n}\)
H <sub>2</sub> O	1368	0.08	$1368 \pm 2.71$
	1000		
$\mathrm{H}_{2}\mathrm{SO}_{4}$	344	0.36	$344 \pm 3.06$
HCl	744	0.33	$744 \pm 6.09$
HNO <sub>3</sub>	808	0.32	808 ± 6.36
CH <sub>3</sub> COOH	424	0.76	$424 \pm 7.98$

#### Salts Effect

Fixing all previous attained experimental parameters whither chemical or physical. The affect of salts solutions as a carrier streams on accumulation of precipitated particles or its dispersed action. In general; it

was noticed that (refer is mode in Fig. 4) a decrease in response profile which might be attributed; that the precipitated particles is dissociated or due to increased solubility. On this basis, distilled water was used as the most suitable solvent for the reaction.

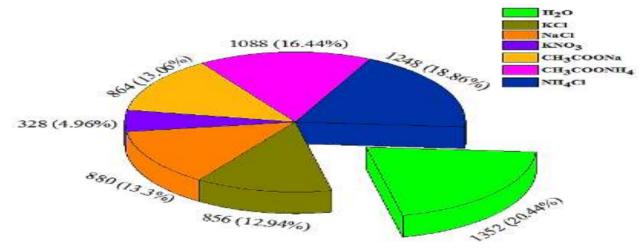


Figure 4: 3Dimintion pie percentage representation of the contribution of each salt solution (B) using Mefimanic acid (5mmol/L) - 8-HQ (7mmol/L) system

## **Physical Parameters**

#### **Electronic Filter Effect**

Using all optimum conditions concerning chemical concentration for two line manifold design system. Studying the effect of RC- low band pass electronic filter variables through the use of 5mmol/L of Mefenamic acid at  $79\mu L$  sample segment.

Electronically noises filter of low band pass with a time constant of 0.1632- 3.974 sec. to overcome the little bit tiny noise developed by the pulse effect of the peristaltic pump. Using this kind of low band pass electronic filter affect on the measured sensitivity, therefore electronic filter was not used and table 3 in which shows the summary of results from this study.

Table 3: Effect of electronic filters on precipitate response expressed as an average peak heights  $\bar{y}_i$  (n=3)

Independent variable of electronic filter response (Sec.)	Dependent variable (ỹi) Average (n=3) diverged light response measured at 0-90° expressed in mV	RSD%	Reliability(two tailed) at 95% confidence level $\bar{y}_i$ (mV) $\pm$ $t_{0.025,2}$ $\sigma_{n-1}$ / $\sqrt{n}$
Without filter	1364	0.07	$1364 \pm 2.73$
0.1632	1346	0.08	$1346 \pm 2.78$
0.3196	1339	0.08	$1339 \pm 2.61$
0.68	1268	0.10	$1268 \pm 3.28$
0.8364	1210	0.12	$1210 \pm 3.53$
1.6728	1187	0.11	$1187 \pm 3.33$
3.974	944	0.16	$944 \pm 3.78$

#### Flow Rate Effect

At the same of optimum parameters in the two line of manifold design for the determination of Mefenamic acid using 8-HQ as a precipitating agent, the variable flow rates ranging (0.4-3.5)ml/min for carrier stream and (0.4-3.6)ml/min for 8-HQ (5mmol/L) line were used. It can be noticed that at low flow rate there is an increase in dispersion and dilution due to the increase of area of precipitate section in flow cell and an increase of  $\Delta tB$  (base width of response).

While an increase of flow rate results in a decrease of sample section; in addition to compact of merged layer due to rashness of successive travelled sample segment zones producing an effect similar to Doppler effect causing an increased of diverged beam of incident light as illustrated in figure 5, and which the optimum flow rat 2.9 & 3 for carrier stream and precipitating agent line respectively which is chosen on the basis of better sensitivity. The results were tabulated in Table 4.

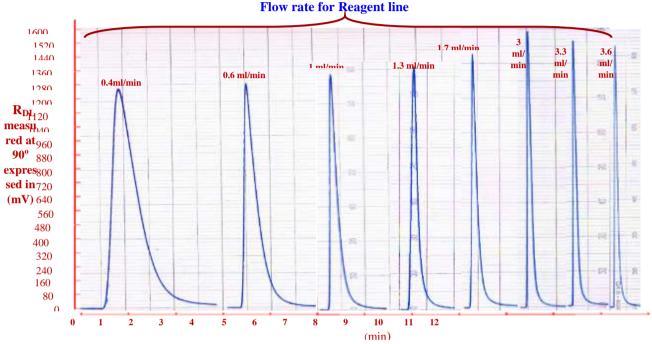


Fig. 5: Effect of flow rate on diverged beam of light vs. time profile

Table 4: Effect of flow rate on the variation of diverged light response and tabulate all available data obtained practically, calculated as obtained by best fit mathematical model, and smoothed digital filtering using Savitzky-Golav data treatment

Indepe	Dep	Dependent variable. Average (n=3) diverged light response measured at 90°													
ndent		expressed in mV													
variabl		Mathemati	Savitzky-												
e of	Flow 1	rate	Average		Reliability(two	$\Delta  ext{t}_{ ext{b}}$	cal model	Golay filter							
pump	(mL/m	nin)	peak	RS	tailed) $\bar{y}_i(mV)\pm$	(Sec)	$\mathbf{\hat{y}}_{\mathbf{i}}$	$\mathbf{\hat{y}}_{i(S-G)}$							
Speed	Line	Line	height(	<b>D</b> %	$\mathbf{t_{0.025,2}} \ \mathbf{\sigma_{n-1}}/\sqrt{\mathbf{n}}$										
	no. 1	no. 2	$\bar{\mathbf{y}}_{\mathbf{i}}$ )												
5	0.4	0.4	1272	0.11	$1272 \pm 3.60$	162	2.239	0.176							
10	0.6	0.6	1304	0.10	$1304 \pm 3.38$	114	2.359	0.167							
15	1	1	1344	0.10	$1344 \pm 3.28$	72	2.479	0.159							
20	1.3	1.3	1368	1.09	$1368 \pm 3.18$	46	2.072	0.191							
25	1.7	1.7	1448	0.08	$1448 \pm 2.93$	34	2.006	0.197							
30	2.9	3	1592	0.07	$1592 \pm 2.81$	30	3.029	0.130							
35	3.2	3.3	1576	0.08	$1576 \pm 3.11$	28	3.112	0.127							
40	3.5	3.6	1520	0.09	$1520 \pm 3.33$	26	3.156	0.125							

 $\Delta t_b$  (sec) : Time lapse for the preciptate response within measuring cell or peak base width Line no.1 = carrier stream (H<sub>2</sub>O), Line no. 2= 8-HQ (5mmol/L)

#### Effect of Sample Volume

The study was carried out using variable sample volume extended from 79-  $329\mu L$ .

All other variable were kept unchanged. It was

noticed the increment in responses as the sample volume is increased reaching  $179\mu L,$  more than; the value an increase of  $\Delta t_B$  with non- returned pen recorder to zero base line due to different particulate formed and difficulty of re-evacuation of whole system

from large molecules formed during commencing the precipitation. Open valve model for discharging the sample plug from injection valve. Table 5 summed up the obtained results.

Table 5: Variation of injected sample volume on diverged light response tabulation all available data obtained practically, calculated as obtained by best fit mathematical model, and smoothed digital filtering using Savitzky-

Golay data treatment

length	Independe	Dependent v	Dependent variable. Average (n=3) diverged light response measured at											
of	nt variable		0- 90o expressed in mV											
sample	sample		Practio	eal lab. Value		Mathemati	Savitzky-							
loop(c	loop	Average	RSD	Reliability(two	$\Delta tb$	cal model	Golay filter							
m)	volume	peak	%	tailed)	(Se	$\mathbf{\hat{y}_{i}}$	ŷi (S-G)							
r=0.5m	$\mu { m L}$	height(ÿi)		$\bar{y}i(mV) \pm t0.025,2$	<b>c</b> )	-								
m		-												
10	79	1590	0.06	$1590 \pm 2.53$	30	1589.997	1944.635							
16.43	129	2320	0.05	$2320 \pm 2.78$	39	2320.469	2118.625							
20.13	158	2500	0.05	$2500 \pm 3.01$	44	2496.024	2380.675							
22.80	179	2560	0.05	$2560 \pm 2.96$	49	2566.660	2677.419							
29.17	229	2920	0.04	$2920 \pm 3.03$	53	2913.249	2924.722							
35.54	279	3180	0.04	$3180 \pm 3.28$	55	3185.317	3091.160							
41.91	329	3220	0.04	$3220 \pm 3.53$	58	3218.284	3175.043							

#### **Purge Time Effect**

This study was conducted using different variable allowed under control time (5- 45 sec) used to purge the plug from injection valve. It was found that increase of peak height with increase of departure time of

sample segment from injection valve up to 40 sec. it was found that no significant difference of using 40 sec to 45 sec could be followed. Therefore, 40 sec was the optimum that can be used to conduct the research at hand. The summary of results tabulated in Table 6.

Table 6: Variation of purge time on diverged light response and tabulate all available data obtained practically, calculated as obtained by best fit mathematical model, and smoothed digital filtering using Savitzky-Golay data treatment

Independent	Dependent v	variable.	Average (n=3) diverged l	ight response me	easured at 90°							
variable of	expressed in mV											
Purge time		Practical	lab. value	Mathematical	Savitzky-							
(Sec.)	Average	RSD%	Reliability(two tailed)	model	Golay filter							
	peak		$\bar{\mathbf{y}}_{i}(\mathbf{mV}) \pm \mathbf{t}_{0.025,2} \; \sigma_{n-1} / \sqrt{\mathbf{n}}$	$\mathbf{\hat{y}_{i}}$	$\mathbf{\hat{y}}_{ ext{i(S-G)}}$							
	$\mathbf{height}(\mathbf{\bar{y}_i})$											
5	520	0.45	$520 \pm 5.84$	520.351	620.478							
10	840	0.26	$840 \pm 5.49$	837.190	831.159							
15	1200	0.21	$1200 \pm 6.31$	1209.833	1226.538							
20	1740	0.11	$1740 \pm 4.92$	1720.333	1717.018							
25	2160	0.07	$2160 \pm 4.41$	2184.584	2141.649							
30	2460	0.05	$2460 \pm 3.06$	2440.333	2418.707							
35	2500	0.05	$2500 \pm 2.96$	2509.834	2540.082							
40	2560	0.04	$2560 \pm 2.58$	2557.190	2566.709							
Open valve	2560	0.04	$2560 \pm 2.68$	2560.351	2559.582							
(45)												

#### **Reaction Coils Effect**

Variable sample loop sizes (157-1301µL) were studied which was attached after Y-junction (refer is made to Fig. 6-A). The reason in why conducting this study is because of having the rearrangement of precipitated particulate or re-crystallization. It was found that; the increase of coil length causes a decreased

sensitivity in general (refer is made to Fig. 6-A&B). This might be explained, that the formation of larger particles is probably the reason by forming a saturation of signal to noise ratio (S/N) output and the difficult associated in the departure of reacted sample too plug due to the enlargement of particulate sample. The condensation of

precipitated particles causes an increase particulate weight and spreading it on a wider surface area and the decrease of particles that reflect and diverge the light. It was formed also in using glass coils; a difficultly is associated with its use while in using teflon tubing an improved signal responses was at hand, due to hydrophobicity the lack of attraction between. So delayed reaction coils were avoided for use.

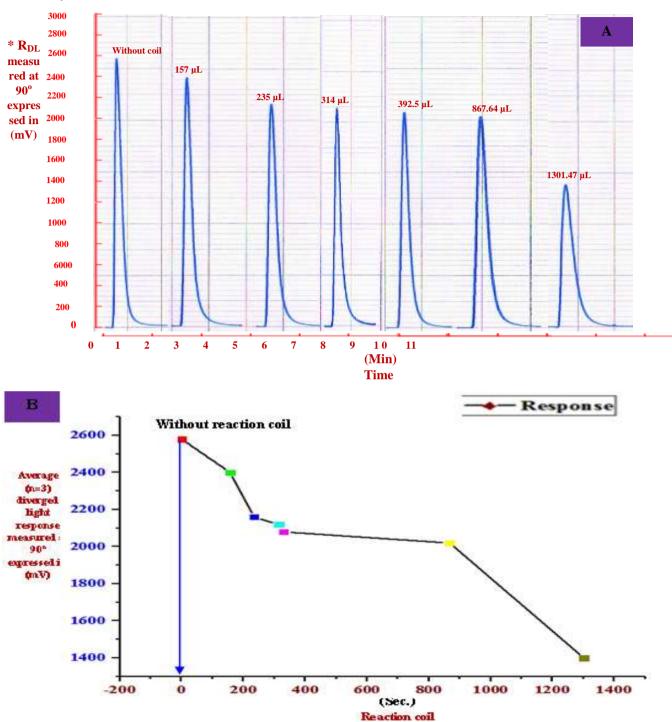


Fig. 6: Effect of different length and material of reaction coil on:-

- Segment precipitate plug that will affect on the whole measurements and profile.
- Laboratory measurements values produced from diverged beam of light using Mefenamic acid (5mmol/L)- 8-HQ- (5mmol/L) system

#### Calibration Graph

Fixing all physical as well as chemical parameters that were studied previously. A series of standard solutions of Mefenamic acid (0.005-7) mmol/L were prepared. The

measurements were conducted using ISNAG-fluorimeter, it was noticed that from the obtained peak height and scatter points plot(fig. 7-A); that best fit extend from 0.005-6 mmol/L. Correlation coefficient of  $\mathbf{r}$  =

**0.9956** and percentage capital **R**-square gave 99.12% (fig. 7-B) was obtained. While dealing with high concentration i.e.; > 7mmol/L decreases of correlation coefficient might be due to accumulation and agglomerate of precipitated particles causes an increase particulate weight which in turn to lead difficulty discharging the particles from flow cell, causes a broadening in peak height and decrease in correlation coefficient value. All results summed up in table 7. The new development methodologies were compared with classical methods of determination using 8-Hydroxyguinoline as a precipitating with optimum concentration

5mmol/L. The extent of linear plot was 0.001-0.09 mmol/L, and in additional classical method (uv-spectrophotometer at 285nm [3] was compared. The linear plot was from 0.001-0.09mmol/L. All the results tabulated in Table 7. Limit of detections were studied for the two used methods. The results are tabulated of the table 7. It can be found that the newly developed methodology was 5 $\mu$ mol/L using successive dilution while the other classical methods was 0.8 $\mu$ mol/L while the repeatability and the trustability for 2 &5 mmol/L having RSD% < 0.2% while classical method < 4% (Table 7).

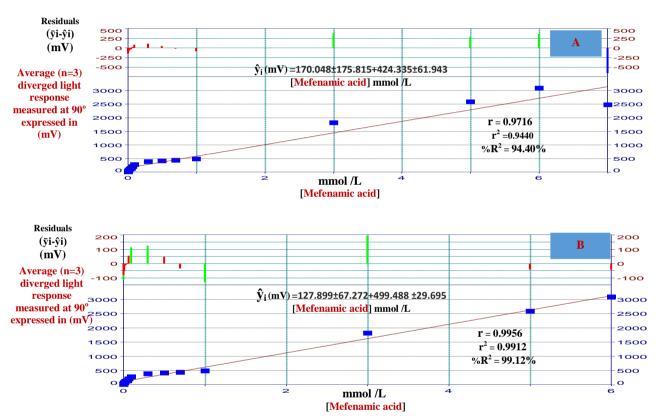


Figure 7: (A) Variation of scattered diverged light at range (0.005-7mmol/L), n=15 against Mefenamic acid concentration (B) Calibration graph deduced from scatter point plot at range (0.005-6mmol/L), n= 14 against Mefemanic acid concentration

Table 7: Summary results of different assessment methods for determination of Mefenamic acid.

Table 7: St	ишш	ary res	uits of ai	nerent assessment	methous	tor ae	ermn	iation of Me	tenamic a	acia.		
		[Mefe	namic	Linear		t- va	lue	Detection	n limit	Rej	peatability at	
Method		a	cid]	regression	r	at 95	%, n-			959	% confidence	
		mr	nol/L	equation	r2	2					level, n-1	
				at 95%, n-2	R2 %							
	(n)			New developed			tca	72			Reliability	
				$\hat{y}i$ (mV)= a± Sat+			1	lu <sub>2</sub>	a)		of	RS
	ıts		a)	b± Sbt [x] mmol				gradual ıum	slope	mmol/L	average	D
	1e1		range	/L					slc	uo.	diverged	
	en		[a]	UV-		n-2		the inin on	on	иu	light	
	ur	ਰ		spectrophotome		_				id] ı		
	of measurements	ured	dynamic	try		ttab t0.025,		on e m rati	based o 3 /slope	acic	ӯi (mV)	
	me	ısı	na	$\hat{y}i = a \pm Sat + b \pm$		ttab t0.02		ed c the ntra	ba 3 / s		Average of	
	of	Meası	dy	Sbt [x] mmol/L		· II		of 1	cal b	amic	absorbance	
		2				ttab		ly b on c	·5 II	ıar	<b>y</b> i	
	No.		Linear			ŧ		tically l dilution co	×ď	[Mefena	Average of	
			<u>:</u> 5					ics ilu	eo	Iei	turbidomet	
			' '					Practically dilutio	The		ry	
								Pre			ӯі (FTU)	
								1			ÿi ±t0.025,	
1	ı	1	1	I	1	1	l	I	1			1

					Turbidometry 0-						n-1 σn-	
					180o						1/√n	
					ŷi (FTU)= a±						n=8	
					Sat+b± Sbt [x]						n 0	
					mmol/L							
					11110172	0.971						
8		15				6	2.160<<					
90		10		_	170.048±	0.944	14.797			2	1364 ±1.321	0.116
olo				0.005- 7	175.815+	0	11			_	1501 -11021	0.110
po	er			100	$424.335 \pm$	94.40						
th	Je1			0.	61.943	%						
ue l	ii.		7		[Mefenamic			(5 µmol/L)	(0.601			
اق	[Or		5-		acid]mmol/L			0.216	µmol/			
be	ÐΓ		0.005-7					μg/sampl	L)			
Newly developed methodology	ISNAG- fluorimeter		0			0.995		e	25.941			
eV	¥	14			105 000 05 050	6	2.1.		ng/	5	2580 ±1.957	0.076
þ	$\mathbf{z}$			0.005-6	127.899± 67.272+	0.991	2.179<<		sampl			
vly				00	$499.488 \pm 29.695$	2	36.652		e			
e,				0.0	[Mefenamic	99.12						
Z					acid]mmol /L	%						
	(6											
	V- absorbance( 3) at λmax =285nm	11				0.995						
	oce 5n			6	0.343± 0.074+	2	2.262<<	$(0.8\mu mol/L)$	(3.544	0.0	$0.999 \pm 0.026$	3.103
	28			0.0	$25.395 \pm 1.873$	0.990	30.671	)	μmol/	3		
n	orb x =			0.001- 0.09	[Mefenamic	5		0.772	L)			
d c	າສ			00	acid]mmol /L	99.05		ug/sample	3.421			
se	ah Yu		_	0.		%			μg/			
ba	UV- at 2		60:						sampl			
ַס	D "		0-						e			
Used method based on			0.001-0.09			0.000						
ne		11	0.			0.998		((0.0	(0.100		490 10 079	0.000
d 1	tr	11		6	9.098± 16.260+	1	2.262<<	((0.8µmol/	(0.103	0.0	436 ±0.953	0.262
$\mathbf{s}_{\mathbf{e}}$	ne (			0.0	8730.313±	0.996		L) 1.544	μmol/	5		
D	II Jo				412.815	1	47.838		L)			
	Turbidometry (NTU)			0.001- 0.09	[Mefenamic	99.61 %		ug/sample	0.199			
	url			0.0	acid]mmol/L	%			μg/			
	Ţ				aciujiiiiioi/L				sampl			
									e			
			l		1				1			

$$\mathbf{t_{tab~(calibration~graph)} = t_{0.025,~n-2}},~ \mathbf{t_{cal}} = \frac{\left| \mathbf{r} \middle| \sqrt{n-2} \right|}{\sqrt{1-\mathbf{r}^2}} \right],~ \mathbf{t_{tab}} = \mathbf{t_{0.025,~n-1=}~t_{0.025,~7=~2.365}},~ \mathbf{r:~correlation~coefficient,~r^2:~coefficient~of}$$

determination&  $R^2\%$ : percentage capital R- square, %RSD = Percentage relative standard deviation.  $\bar{y}_i$  = practical value for (n=8) in mV for newly developed method, FTU for turbidometry method, and without unite for spectrophotometry method,  $\hat{y}_i$  =estimated value in mV for newly developed method (volume 179 $\mu$ L) FTU for turbidometry method (volume of 8ml), and without unite for spectrophotometry method (volume of 4ml),

#### **Application**

The continuous flow injection analysis via diverged light response using ISNAGfluorimeter achieved in this work was used for the analysis of Mefenamic acid in the four different drug manufactures (Ponstan-forte (German)- 500 mg, Ponstan-forte (Egypt)-500 Merfile-500 (India)-500 mg, mg, ponstidin- 500 (Iraq)- 500 mg) and was compared with two methods which includes UV-spectrophotometric via the measurement of absorbance at  $\lambda_{max} = 285$ nm by UVspectrophotometer,[3] (UV-1800 Shimadzu), turbidometry via Turbidity-meter, HANNA, (Hungary).

The measurement of scattered light at 0-180° for yellow precipitate particles of Mefenamic acid-8-HQ (5mmol/L) system. A series of solutions were prepared of each drug (5mmol/L) by transferring 1ml to each five

volumetric flask (10ml), followed by the addition of gradual volumes of standard solution of Mefenamic acid (0, 0.8, 1, 3, and 5ml) of 5 mmol/L to obtain (0, 0.4, 0.5, 1.5, mmol/L) when use ISNAGfluorimeter (newly developed methodology), in UV-spectrophotometer method transferring 0.06 ml from 5mmol/L sample solution to each five volumetric flask, followed by the addition of gradual volume of standard solution of Mefenamic acid (5 mmol/L) (0, 0.01, 0.02, 0.03, and 0.04ml) to obtain (0, 0.005, 0.01, 0.015, and 0.02 mmol/L), in addition to Turbidomtric method that depend on the measurement at 0-180°. The series of solutions were prepared by transferring 0.06 ml of 5 mmol concentration of each sample, followed by the gradual addition of 0, 0.01, 0.02, 0.03, and 0.04 ml from standard solution of Mefenamic acid (5mmol/L) to obtain 0, 0.005, 0.01, 0.015, and 0.02 mmol/L.

The measurements were conducted by three methods. Fig. 8-A, B, C and D shows standard addition calibration graphs using

newly developed methodology. The results were summed in table 8 at confidence level 95% (2-tailed), showing practically content of Mefenamic acid in each sample of drug using three different methods and efficiency of determination.

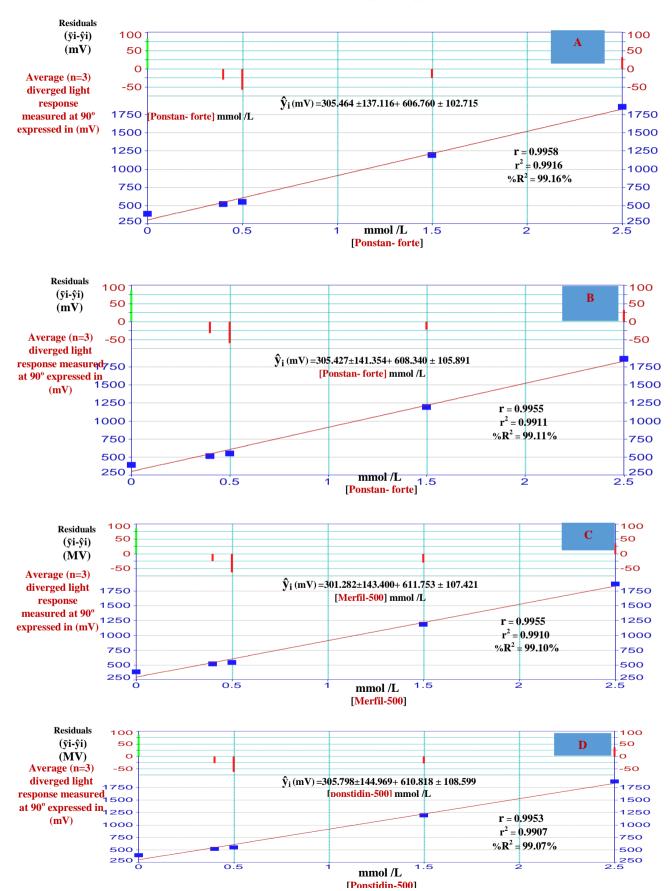


Fig. 8: Standard addition calibration graph using ISNAG-fluorimeter for: A - Ponstan-forte (German), B - Ponstan-

forte (Egypt), C- Merfil-500 (India), D- ponstidin-500 (Iraq) Residual = ( $\hat{y}i-\hat{y}i$ ) in mV,  $\hat{y}i$  = practical value,  $\hat{y}i$  = estimated value

Table 8: Summary of results by standard additions method for the determination of Mefenamic acid by different

met	hods			<i>9</i> 00		AG flu				n mV						<u> </u>
	Α		ient	645 g)	V-spe	ctrome	ter ȳ <sub>i</sub>	(n=3)								
	ıntr		gred	0.120645 wi (g)	Tur	bidome	eter ÿ <sub>i</sub>	(n=3) i	n FTU							
	Name, coted value, company And country		ive ing		Vol.	ume of	f Mefe	namic	acid	$\hat{\mathbf{y}}_{i} = \mathbf{a} \pm \mathbf{S}_{a}\mathbf{t} + \mathbf{b} \pm \mathbf{s}$	$\mathbf{r}$ $\mathbf{r}^2$	Practactiv		conte	ent of	
	any A		at 95%		[Me	fenami	c acid]	mmol/	L	S <sub>b</sub> t [Sample] mmol/L	R <sup>2</sup>	Conc		_Weig ht in		
	comp	Confide	t for t	Sample weight equivalent to (5mmol/L of the active ingredient.	0.0	0.8	1	3	5	at confidenc		prep sample	ared e in	1		
nple	alue,	interva for the average		ght enet	0.0	0.4	0.5	1.5	2.5	e level 95%, n-2		10 ml	100 ml	Wi (g) ±t <sub>0.025</sub> , <sub>n-1</sub> σ <sub>n-</sub>	n/√n	rmins
Number of sample	oted v	average weight o	$\frac{1}{2}$ coretical conterm (mg)±1.96 o <sub>n-1</sub> / $\sqrt{n}$	weig L of tl	0.0	0.01	0.02	0.03	0.04					$\frac{1}{\sqrt{n}}$	, n-1 <b>G</b> p	f dete
$_{ m mber}$	me, cc	tablets Wi (g)	Theoretical WI (mg)±1.96	nple nmol/]	0.0	0.00 5	0.01	0.01 5	0.02					95%	ets ±t <sub>0.025</sub>	ncy of
Nu	Naı	$1.96 \sigma_{\rm n}$	ĀŢ.	Sar (5n	ÿi f	or n= 3									In tablets $\overline{W}$ i (mg) $\pm t_{0.025, n-1} \sigma_{n-1} / \sqrt{n}$	Efficiency of determination
1		$0.734 \\ 5\pm \\ 0.00 \\ 38$	500 ± 2.58 7	0.177 23	38 8	518	550	119	185	305.464 ±137.116+ 606.670 ± 102.715 [Ponstan- forte] mmol/L	0.99 58 9916 99.1 6%	0.50	5. 035 1	0. 1214 9 ± 0.001 99	503.4 96 ± 8.247	100. 70%
					1.0 28	1.14	1.31	1.48	1.69	0.998 ±0.073+ 33.260 ± 6.062 [Ponstan- forte] mmol/L	.995 1 .990 3 9.03	0.03	5. 003 0	0.120 72 ± 0.002 09	500.2 87± 8.662	100.0 6%
	Ponstan-forte 500mg- German				258. 46	304.8 6	348. 18	400. 87	429. 11	260.834 ±16.107+ 8746.200 ± 1315.213 [Ponstan- forte] mmol/L	.996 7 .993 4 9.34 %	0.02 98	4. 970 4	0.119 93 ± 0.001 84	497.0 29 ± 7.626	99.41
2		0.729 6± 0.00 14		0.1760 5	39 1	516	549	119	185 8	305.427±14 1.354+ 608.340 ± 105.891 [Ponstan- forte] mmol/L	0.99 55 0.99 11 99.1 1%	0.50 21	5. 020 1	0.121 14 ± 0.001 6	502.0 66 ± 6.631	100. 41%
					1.0 21	1.15	1.31 9	1.49	1.69	0.998 ±0.061+ 33.720 ± 4.824 [Ponstan- forte] mmol/L	.997 .994 9.40	0.02 96	4. 930 8	0.118 98 ± 0.001 81	493.0 80 ± 7.501	98.6 2%
	Ponstan-forte 500mg- Egypt				25 6.9 3	308. 11	346. 29	408.	428. 44	260.972±18 .631+ 8724.40 ± 1521.200 [Ponstan- forte] mmol/L	.995 5 .991 1 9.11	0.02	4. 985 5	0.120 30 ±0.00 169	498.5 48 ± 7.004	99.7
3	Merfil- 1 500				38 6	520	544	118 8	186 6	301.282±14 3.400+ 611.753 ± 107.421	.995 5 .990 9	0.49 25	4. 924 9	0.118 83 ±0.00 149	492.4 81 ± 6.175	8.50%

		0.040	<b>T</b> 00	0.0000						F3 /F (#11	0.00					
		0.840	500	0.2028							9.09					
		8±	±	8						mmol /L	%					
		0.00	2.67		1.0	1.16	1.31	1.48	1.69	0.998±0.05	.997	0.02	4.	0.119	495.8	99.1
		48	6		13	5	4	7	1.03	1+ 33.560 ±	7	96	958	64 ±	20±	6%
					10	o o	4	′	1	4.175		90	3	0.001	6.921	0 / 0
											.995		3	67	6.921	
										[Merfil]	4			67		
										mmol /L	9.54					
											%					
																0 4 704
					25	306.	346.	402.	431.	260.462±17	.996	0.02	4.	0.118		8.15%
					8.0	47	19	81	06	.402+	2	94	907	41 ±	26 ±	
					8					$8846.00 \pm$	.992		3	0.001	8.247	
										1420868	4			99		
										[Merfil]	9.24					
										mmol/L	%					
					39	522	548	119	186	$305.798 \pm 14$	.995	0.50	<b>5</b> .	0.120		100.1
					2			2	8	4.969+	3	06	006	80 ±	30 ±	3%
4										$610.818  \pm$	.990		4	0.001	7.626	
4										108.599	7			84		
		0.598	500	0.1444						[ponstidin						
		7±	±	6						mmol/L	9.07					
		0.00	1.92							1 mmor/L	%					
		23	1		1.0	1.16	1.31	1.48	1.69	0.999±0.06	.996	0.02	4.	0.119	495.7	9.15%
					19	0	8	2	8	$4+33.600 \pm$	6	97	957	62 ±	$27 \pm$	
										5.075	.993		3	0.019	8.123	
										[ponstidin	.995 3			6		
										**	_					
										] mmol/L	9.33					
											%					
	0				25	310.	344.	401.	430.	259.924±19	.995	0.02	4.	0.118	490.3	8.07%
	50( g				4.8	08	89	44	07	.945+	0	94	903	31 ±	34±	
	n-{ ra(				8	00	00	11	· ·	8834.800 ±		71	4	0.001	7.004	
	Ponstidin-500 500mg- Iraq									1628.567	.990		-1	69	7.004	
	sti ng										0			00		
	on On									[ponstidin	9.00					
	Ρ. 50									] mmol/L	%					
	Б			L .	/ TD		CALAG	m ·		• • • • • • • • • • • • • • • • • • • •	• • •	T 7 7		, .		

 $\hat{y}_i$ : Estimated response value (mV) for ISNAG- fluorimeter, without unite for UV-spectrometric method, and Turbidomtric method (FTU)) for (n=3), [sample]: drug concentration (mmol/L), r: correlation coefficient, r<sup>2</sup>: coefficient of determination& R<sup>2</sup>%: percentage capital R- square,  $t_{0.025, \infty} = 1.96$  at 95%,  $t_{0.025, 2} = 4.303$  for n-1  $t_{0.025, 3} = 3.182$ . For n-2 drawing volume of 1ml for ISNAG-fluorimeter and 0.06 for two classical methods

#### Conclusion

The newly developed adopted methodology in this research work was put into a paired t-test for the sake of accepting it as an alternative method for analysis and assessment of Mefemanic acid with standard used method. Mainly British Pharmacopoeia (B.P) [31], and turbidometry method (Scheme 2), or rejecting it as an alternative method. The assessment is made on how much they are correlated as a methods and if there is any significant difference that will work against the developed method.

On this basis three assumptions statistically is made [32, 33]. There is no significant difference between the means of all used four methods (i.e.; undistinguishable differences between the method) and if  $\mu$  indicates the mean then it will annotated with specified term representing the method used as such

# H<sub>o</sub> = Null hypothesis = No significant difference between

H<sub>o</sub>:  $\mu_{B.P} = \mu_{ISNAG-fluorimeter} = \mu_{Turbidometry} = \mu_{uv-spectrophotometry}$ 

 $H_1$  (alternative hypothesis):  $\mu_{B,P} \neq \mu_{ISNAG-fluorimeter} \neq \mu_{Turbidometry} \neq \mu_{uv\text{-spectrophotometry}}$ .

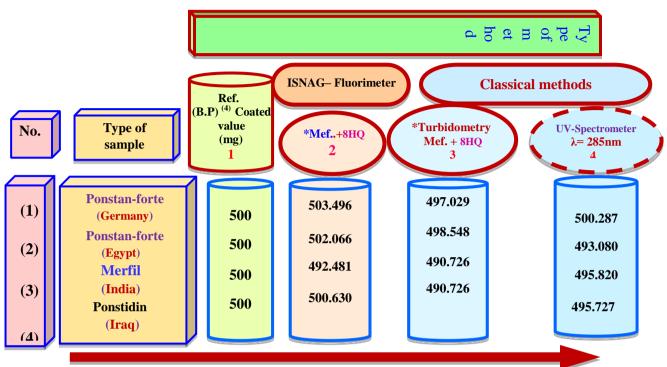
Conducting paired t- test will all possible pairs (i.e.; 5- pairs) five that are as follows: Vs. ISNAGfluorimeter British Pharmacopoeia (BP), ISNAG Vs. turbidometry, ISNAG Vs. spectrophotometry, spectrophotometry Vs BP and turbidometry Vs BP. As ISNAG- fluorimeter being the suggested alternative or equivalent method of assessment of the drug which challenges the available official method as ISNAGfluorimeter as an instrument is new in its whole properties of working and presenting results for determination.

So therefore, it is the one who is its capability is under question and its approval as a method with the existing method and the used ones. Following table 9, it can be found that there are five comparisons. As it compare, ISNAG- fluorimeter method with the other there standard method as shown above. Which significance test indicate that at 95% confidence ( $\alpha = 0.05/2$  two tailed) there is no significant difference between the

newly developed method and standard methods.

Therefore, the analyst should be able to choose any method for analysis i.e.; ISNAG-

fluorimeter or spectrophotometry or turbidometry. Thus accepting null hypothesis. This indicate that the high efficiency of ISNAG- fluorimeter as a reliable instrument for analysis of Mefemanic acid.



Path for comparison between four methods in addition to quoted value using ANOVA (Analysis of variance) - one way (F- test)

Scheme 2: Summed up the results for three different methods in addition to quoted value and four different samples for ANOVA \* ISNAG- fluorimeter (8HQ: ISNAG-fluorimeter using Mefenamic acid- 8HQ (5mmol/L) system. \*Turbidometry: Mefenamic acid-8HQ (0.005mmol/L) system

Table 9: Paired t-test for the comparison between four different methods of four samples for the analysis of Mefenamic acid in drugs for n=4 at 95% confidence level ( $\alpha=0.05$ ) and DF = 3

Therefore the transfer of the		Paired diffe				
Paired		Standard	$\mathbf{t_{cal}}$	$\mathbf{t_{tab}}$	Significant	
	Xd	Deviation			(2 tailed)	
		$(\sigma_{n-1})$				
Pair- 1	- 0.332	4.932	- 0.13	5 < 3.182	0.902 > 0.05	
ISNAG fluorimeter - BP					Not significant	
Pair- 2	3.440	5.129	1.341	< 3.182	0.272 > 0.05	
ISNAG fluorimeter- spectrophotometry.					Not significant	
Pair- 3	5.411	3.571	3.030	7< 3.182	0.0563 > 0.05	
ISNAG fluorimeter-Turbidometry.					Not significant	
D : 4	9.779	9.000	9.594	z 0 100	0.0050 > 0.05	
Pair- 4	3.772	2.989	2.524	< 3.182	0.0859 > 0.05	
BP - spectrophotometry					Not significant	
Pair-5	5.743	4.124	2.785	< 3.182	0.069 > 0.05	
BP - Turbidometry					Not significant	

DF: Degree of freedom (n-1) = 3, Xd: average of difference between two methods,  $t_{tab} = t_{0.025, 3} = 3.182$ ,  $tcal = Xd \sqrt{n} / \sigma n - 1$  at 95 %

#### References

- 1. Neuvonen P, Kivisto K (1994) Enhancement of drug absorption by antacids. An unrecognized drug interaction. Clin Pharmacokinet, 27: 120-128.
- 2. Tall A, Mistilits S (1975) Studies on Ponstan (Mefenamic acid): I. Gastrointestinal bloodloss; II. Absorption and excretion of a new formulation. J. Int. Med Res (UK)., 3 (3): 176-182.
- 3. Harinder S, Rajnish K, Pinderrjit S (2011) "development of uv-spectrometric method for estimation of Mefenamic acid in bulk and pharmaceutical dosage forms" journal International of Pharmacy and Pharmaceutical Sciences, 3 (2): 0975-1491.
- Glazko A (1966) Experimental observations of flufenamic, Mefenamic, and meclofenamic acids. Part III. Metabolic disposition, in Fenamates in

- Medicine. A Symposium, London,. Annals of Physical Medicine, Supplement, 23-36.
- 5. Budoff P (1979) Use of mefenamic acid in the treatment of primary dysmenorrhea. JAMA, 241: 2713-2716
- 6. Buchanan R E (1974) The breast milk excretion of mefenamic acid. Curr. Ther. Res., 10: 592.
- 7. Champion G, Graham G (1978) Pharmacokinetics of non-steroidal antiinflammatory agents. Aust. NZ. J. Med., 8 (1): 94-100,
- 8. Al-Abdaly ZZ (2005) Spectrophotometeric Determination of p-Aminobenzoic Acid-Application to Pharmaceutical Preparations, M.Sc., Hesis, Mosul University, 61.
- 9. Albero M, Sanchez C, Farcia M (1995) Flow-injection Spectrofluorometric Determination of Flufenamic and Mefenamic, J. Pharm. Biomed .Anal.,13: 1113-1117.
- 10. Al-Irhayim, A (2004) Spectrophotomtric Assay of Isoniazide in Tablet, M.Sc., Thesis, Mosul University, 33.
- 11. Aman T, Asrar A, Mateen B (2005) Colorimetric Determination of Two Nonsteroidal Anti-inflammatory Drugs Using p-dimethylaminocinnamaldehyde, Anal. let., 38: 1899-1912.
- 12. Chilukuri S, Ambati R (1986) Spectrophotometric Determination of some Analgesic and Anti-inflammatory Agents with Methyl-2-Denzothiazolinone Hydrazone Hydrochloride, Mikrochimi.a Acta, 97: 237-244.
- 13. Dinc E, Yucesoy C, Onur F (2002) Simultaneous Spectrophotometric Determination of Mefenamic Acid and Paracetamol in a Pharmaceutical Preparation Using Ratio Spectra Spectrophotometry Derivative and Chemometric Methods. Pharma. J. Biomed. Anal., Vol.28, pp. 1091-1100.
- 14. EL-Sherif Z, Walash M, EL-Tarras M, Osman A (1997) Colorimetric Determination of Two Nonsteroidal Anti-inflammatory Drugs Using p dimethylaminocinnamaldehyde, Anal. lett., 30:1881-1896.
- 15. Fatma A, Aly Salma, A AL-Tamimi, Abdulrahman A, ALwarthan (2000) Determination of Flufenamic Acid and

- Mefenamic Acid in Pharmaceutical Preparations and Biological Fluids Using Flow Injection Analysis with Tris (2,2'-bipyridyl) Ruthenium (II) Chemiluminescence Detection, Anal. Chim. Acta, 416: 87-96.
- 16. Garg G, Sarsf S (2007) Simultaneous Estimation of Mefenamic Acid and Ethamsylate in Tablets. Indian J. Pharm. Sci., 69: 2-7.
- 17. Idowut S, Adegoke A, Olaniyi A (2003) Novel Colorimetric Assay of Mefenamic Acid Using 4-Amino-3,5-Dinitrobenzoic Acid (ADBA), Tropical J. Pharm. Research, 1: 15-22.
- 18. Ishidaka O, Shinohara T, Tanaka T, Momose A (1986) Determination of Mefenamic acid in Horse Plasma by High Performance Liquid Chromatography, Japan analyst, 35: 332-334.
- 19. Jaime ND, William AR, Wilson Gisvold's (1991) Text Book of Organic Medicinal and Pharmaceutical Chemistry, 9th Edn., J.B. Lippincott Company, London, 201.
- 20. Idowut S, Adegoke A, Olaniyi A (2003) Novel Colorimetric Assay of Mefenamic Acid Using 4-Amino-3,5-Dinitrobenzoic Acid (ADBA), Tropical J. Pharm. Research, 1: 15-22.
- 21. Chilukuri S, Ambati R (1986)
  Spectrophotometric Determination of some
  Analgesic and Anti-inflammatory Agents
  with Methyl-2-Denzothiazolinone
  Hydrazone Hydrochloride, Mikrochimi.a
  Acta, 97: 237-244.
- 22. Sastry C, Rao A (1987) Spectrophotometric Method for Determination of Pentazocine and Mefenamic acid, Indian J. Pharm. Sci., 49: 95-96.
- 23. Aman T, Asrar A, Mateen B (2005) Colorimetric Determination of Two Nonsteroidal Anti-inflammatory Drugs Using p-dimethylaminocinnamaldehyde Anal. let., 38: 1899-1912.
- 24. EL-Sherif Z, Walash M, EL-Tarras M, Osman A (1997) Colorimetric Determination of Two Nonsteroidal Anti-inflammatory Drugs Using p dimethylaminocinnamaldehyde, Anal.lett., 30: 1881-1896.
- 25. Tabrizi AB (2006) Determination of Mefenamic Acid in Human Urine by Means of two Spectroscopic Methods by

- Using Cloud Point Extraction Methodology as a Tool for Treatment of Samples, Bull. Korean Chem. Soc., 27: 1780-1784.
- 26. Zommer S, Bojarowicz H (1986) Spectrofluorimetric Investigations on Protolytic Equilibria of Mefenamic Acid and Determination by Means Fe (III) in Methanol- Aqueous Media, J. Pharm. Biomed. Anal., 4: 475-481.
- 27. Dinc E, Yucesov C, Onur F (2002) Simultaneous Spectrophotometric Determination of Mefenamic Acid and Paracetamol in a Pharmaceutical Preparation Using Ratio Spectra Spectrophotometry Derivative and Chemometric Methods. J. Pharma. Biomed. Anal., 28: 1091-1100.
- 28. Garg G, Sarsf S (2007) Simultaneous Estimation of Mefenamic Acid and

- Ethamsylate in Tablets. Indian J. Pharm. Sci., 69: 2-7.
- 29. Robert CA, Francis AC (1997) organic chemistry, 6th Ed., Mc G raw-Hill companies, New York, 436.
- 30. Smith JG (2006) Organic chemistry, McGraw Hill, 1st edn., New York, 1003.
- 31. The British Pharmacopoeia Commission Secretariat (2009) Part of the Medicines and Healthcare products Regulatory Agency (MHRA). British Pharmacopoeia, Her Majesty's Stationery Office, London, UK.
- 32. Jeffery JH, Bassett J, Mendham J, Denney RC (1997) Vogel's textbook of quantitative chemical analysis, 5thEd., New York.
- 33. JM Miller, JC Miller (2005) "Statistical and chemometric for analytical chemistry", 5thed, person education limited.