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**RESEARCH ARTICLE** 

# Effects Cuminum cyminum Extracts in Lipid Profile Levels and Internal Tissues of Infected Balb/c Mice with Hydatid Cysts

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#### Abstract

Hydatidosis is one of the most cosmopolitan zoonotic disease that may causes death and still very common in the developing world. The present study aimed to detect the effects of Albendazole and C. cyminum extracts on lipid profile, liver and spleen of infected Balb/c mice with hydatidosis. This study showed positive results for tannins, phenolic compounds, flavonoids and glycosides in alcoholic and aqueous extracts of C. cyminum, while negative result for alkaloids. Significant (p  $\leq$  0.05) increasing of total cholesterol was detected for the third (273.57 mg/dl) and fourth (281.73 mg/dl) positive control Triglycerides shows significant increasing in groups 3 (248.65 mg/dl) and 4 (260.30 mg/dl) of positive control groups. About HDL-C, significant decreasing was recorded in the third and fourth positive control groups (40.87 mg/dl and 33.42 mg/dl, respectively), while significant (p  $\leq$  0.05) elevation was occurred for HDL-C in Albendazole treatment group (60.87 mg/dl) as compared with the negative control group, with no significant differences were reported for HDL-C in alcohol and aqueous extract treated groups. For histological study, the present study showed degeneration of hepatocytes, thickening wall of hepatic blood vessels and infiltration of lymphocytes for Albendazole treatment group, while alcoholic and aqueous extract treated groups showed central vein, hepatocytes and sinusoids with normal sizes. Concerning spleen, results in the current study showed normal shape and size for white pulp, red pulp and central artery of all treatment groups, while edema and necrotic materials appeared in some of positive control groups.

**Keywords:** Hydatidosis, Hydatid cyst, Cuminum cyminum extract Histological effects.

#### Introduction

Echinococcus granulosus is a tapeworm belong family Taeniidae that causes hydatidosis or cystic echinococcosis in human and most of herbivorous animals [1]. The life cycle of *E. granulosus* needs two mammalian hosts, including an intermediate host of herbivorous animal and definitive canine host (dogs), while human is considered as an accidental host [2].

Hydatidosis is one of the most cosmopolitan zoonotic disease that may causes death and still very common in the developing world [3]. The internal organs especially liver and lungs of humans and all intermediate hosts are the main sites for hydatid cyst, which appears like unilocular fluid-filled bladder [4]. Cystic echinococcosis causes pathological damages or dysfunction especially by the gradual effecting of space-occupying repression or displacement of vital host

tissue, vessels or organs [5]. The symptoms depend on the habitat, number and size of the cysts and the complications caused by rupture of cysts [6]. Albendazole is a useful drug for the treatment of hydatidosis [7]. In the last years, some of medical plants are used as alternative treatment instead of the artificial chemical drugs, because it has active anti-microbial properties due to existence of active compounds.

The active compounds are produced due to secondary metabolism in the plants, these compounds including flavonoids, glycosides and phenolic compounds [8]. Cuminum cyminum is one of medical plants belong family Aplaceae that using for digestive system disorders, intestinal helminths eviction and sterilizing of urinary tract [9]. The present study aimed to detect the effects of Albendazole and C. cyminum extracts

(alcoholic and aqueous) on lipid profile, liver and spleen of infected Balb/c mice with hydatidosis.

#### **Materials and Methods**

#### **Plant Extracts**

C. cyminum was obtained from College of Agriculture, Tikrit University, Iraq and it was identified by the National Herbarium of Iraq, Ministry of Agriculture Add paragraph about.

#### **Plant Extracts Preparation**

Mice were obtained from National Center for Drug Control and Research (NCDCR) in Baghdad, Iraq. This study was performed from February of 2016 to June of 2016

# **Phytochemicals Constituents**

Alcoholic and aqueous extracts ofC. cyminum, Dragendroff's test was used to detect Alkaloids, and appearance of red sediment means a positive result [19, 20]. Tannins and phenolic compounds were determined by ferric chloride test, dark blue and blackish green colors are the evidences for the positive result [21], while appearance of black or blackish red color when ferric chloride test used that the evidence for existing flavonoids [22]. Sodium hydroxide test was used to detect glycosides and yellow color produces for the positive result [23].

#### Isolation of *E. granulosus* Protoscoleces

Protoscoleces of *E. granulosus* were isolated from ovine fertile hydatid cysts and conserved it in the 1:4 (v/v) of hydatid cyst fluid and Krep's Ringer solution According to [10]. Krep's Ringer solution was prepared Based on [11]. Viability and number of protoscoleces were done according to [12] using 0.1% of Eosin stain.

#### **Experimental Design**

A total of 40 Balb/c mice with age 12-14 weeks and weight 30-35 gm were divided in to 8 groups included 5 mice per group:

- Negative control group that injected with phosphate buffer saline (PBS) in the peritoneal cavity [13, 14]. PBS was prepared according to [15] during the use.
- Positive control groups (4) which injected with 2500 protoscoleces that suspended in 1 ml of PBS by syringe 3 ml and needle 23G, then dissected and examined after 1, 2, 4 and 6 weeks.
- Treatment groups three groups were infected experimentally first group treated with 15 mg/ kg / body weight daily of Albendazole [16]. Second and third groups were dosed with 500 mg/ kg / body weight daily of alcoholic extract of *C. cyminum* that prepared based on [17] and aqueous extract of *C. cyminum*, which prepared according to [18], respectively. Treatment was continued for 6 weeks, Iraq All.

## **Lipid Profile**

In the present study, total serum cholesterol, serum triglycerides and heavy density lipoprotein-C (HDL-C) were estimated using three commercial kits (Randox, UK) according to the manufacturer's instructions.

#### **Histological Tests**

Histological sections of mice liver and spleen were prepared according to [24] from all mice groups in the present study.

#### Statistical Analysis

Duncan's Multiple Range Test (DMRT) was used to analyze the data statistically with SPSS software program [25].

#### **Results and Discussion**

# Active Compounds of C. cyminum Extracts

The present study showed positive results for tannins, phenolic compounds, flavonoids and glycosides in alcoholic and aqueous extracts of *C. cyminum*, while negative result for alkaloids (Table 1).

Table 1: Active compounds in alcoholic and aqueous extracts of C. cyminum

Activo commonada	Type of extract		
Active compounds	Alcoholic extract	Aqueous extract	
Tannins	+	+	
Phenolic compounds	+	+	
Flavonoids	+	+	
Glycosides	+	+	
Alkaloids	-	•	

The current study agreed with study of [26] that detected tannins and glycosides in C. cyminum extracts and no alkaloids were found. Also, our study agreed with [27] that found flavonoids and glycosides in C. cyminum extracts. The present study agreed with study of [28] for existence of tannins. phenolic compounds, flavonoids and glycosides in *C. cyminum* extracts. chemical detection for active compounds in C. cyminum extracts showed antioxidant activity, enzymatic control, growth inhibition for many organisms included parasites, fungi and bacteria [29, 30, 31]. These active compounds have growth inhibition activity for many parasites that involved granulosus [29, 32].

## **Lipid Profile**

Results for total serum cholesterol, serum triglycerides and heavy density lipoprotein-C (HDL-C) of the present study were showed in Table 2.

#### **Total Serum Cholesterol**

Non-significant variances were recorded for total serum cholesterol between first and second groups of positive control (236.94 mg/dl and 222.29 mg/dl, respectively) with the negative control group 203.30 mg/dl, while, significant (p  $\leq$  0.05) increasing was detected for the third (273.57 mg/dl) and fourth (281.73 mg/dl) positive control groups when compared with negative control.

Also. non-significant differences were reported for total serum cholesterol among all treatment groups (Albendazole 219.34 mg/dl, alcoholic extract 245.74 mg/dl and aqueous extract 237.07 mg/dl) as compared with the negative control group. The rising of total serum cholesterol might due to decrease HDL-C that responsible transporting of cholesterol from somatic cells to hepatic via blood, therefor cholesterol accumulated in the blood [33].

#### **Triglycerides**

For positive control groups, non-significant variances were recorded between groups 1 and 2 (194.65 and 203.90 mg/dl, respectively),

while significant increasing for groups 3 (248.65 mg/dl) and 4 (260.30 mg/dl) when compared with the negative control group (178.00 mg/dl). In addition, there are no significant variances for triglycerides among whole treatment groups; Albendazole treatment group, alcoholic and aqueous extract treated groups (169.54 mg/dl, 198.44 mg/dl and 176.67 mg/dl, respectively) as comparison with the negative control group.

These results due to existence of phenolic compounds, flavonoids and glycosides, which inhabit the reactions of free radicals formation via removing these radicals via antioxidant activity, increasing of antioxidant hepatic enzymes activity such as catalase and superoxide dismutase [34]. In addition, these active compounds increase insulin sensitivity that increases lipoprotein lipase activity, the last can be analyzing triglycerides to fatty acids and glycerol [35].

#### Heavy Density Lipoprotein-C (HDL-C)

In the present study, the positive control groups showed non-significant variances in the first (48.02 mg/dl) and second (49.82 mg/dl) groups, while significant decreasing was happened in the third and fourth positive control groups (40.87 mg/dl and 33.42 mg/dl, respectively) as comparison with the negative control group (47.63 mg/dl). For treatment groups, significant (p  $\leq 0.05$ ) increasing was occurred for HDL-C in Albendazole treatment group (60.87 mg/dl) as compared with the negative control group, while no significant differences were reported for HDL-C in alcohol and aqueous extract treated groups when compared with the negative control group.

Many reasons for HDL-C decreasing: hepatic lipase effected by metabolic disorders, which may cause by parasite and lead to break HDL-C molecule, then take cholesterol from hepatocytes for transforming it to bile salts. Another may explains HDL-C reason decreasing, the lower activity of lysis for very density lipoprotein (VLDL) functional changes in liver that caused by hydatidosis [36, 37].

Table 2: lipid profile of treatment, positive control and negative control groups

Parameters	Total Cholesterol	Triglycerides	HDL-C
Groups	Mean± SD (mg/dl)	Mean± SD (mg/dl)	Mean± SD (mg/dl)
Albend. Treat.	$219.34 \pm 95.83$	$169.54 \pm 29.27$	$60.87 \pm 21.73$
	AB	C	A
Alco. Ext. Treat.	$245.74 \pm 61.49$	$198.44 \pm 57.88$	$42.86 \pm 17.78$
	AB.	$\mathbf{C}$	BC
Aque. Ext. Treat.	$237.07 \pm 55.81$	$176.67 \pm 60.67$	$41.28 \pm 12.53$
	AB	C	BC

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+ve Ctrl. 1	$236.94 \pm 65.08$	$194.65 \pm 39.53$	$48.02 \pm 10.16$
	AB	C	AB
+ve Ctrl. 2	$222.29 \pm 58.72$	$203.90 \pm 30.20$	$49.82 \pm 9.81$
	AB	BC	AB
+ve Ctrl. 3	$273.57 \pm 68.30$	$248.65 \pm 39.68$	$40.78 \pm 9.95$
	A	AB	BC
+ve Ctrl. 4	$281.73 \pm 88.31$	$260.30 \pm 58.93$	$33.42 \pm 5.79$
	A	A	C
-ve Ctrl.	$203.30 \pm 40.62$	$178.00 \pm 17.22$	$47.63 \pm 8.87$
	В	C	В

#### Histological study

#### Liver

The present study showed degeneration of hepatocytes, thickening wall of blood vessels and infiltration of lymphocytes for Albendazole treatment group (Figure 1).

These changes may be relating to immunity and presence of hepatic hydatid cyst. In addition, Albendazole causes infiltration for immune cells [38, 39], while the toxic substances that secreted by parasite cause degeneration through autolysis of hepatocytes [40].

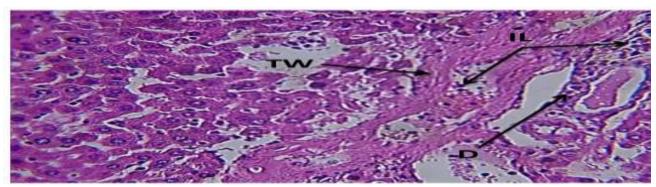


Figure 1: Hepatic section of Albendazole treatment group shows degeneration of hepatocytes (D), thickening wall of blood vessels (TW) and infiltration of lymphocytes (IL), H and E400x

For alcoholic and aqueous extract treated groups (Figures 2 and 3, respectively), hepatic sections showed central vein, hepatocytes and sinusoids with normal sizes. The reason for positive effect of *C. cyminum* extracts on hepatic tissue is presence of

antioxidant compounds such as flavonoids and phenolic compounds that protect liver from parasitic toxins, anti-lipid peroxidation and inhibit lysis of hepatocytes membrane [41, 42, 43].

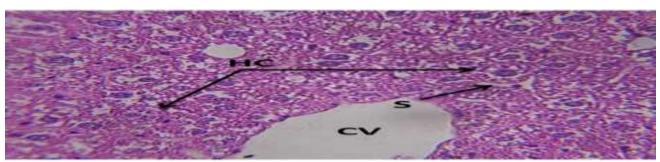


Figure 2: Hepatic section of alcoholic group of *C. cyminum* shows central vein (CV), hepatocytes (HC) and sinusoids (S) with normal sizes, H and E 400 xs

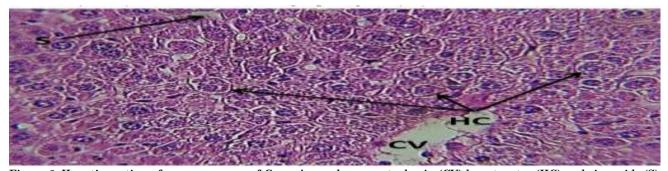


Figure 3: Hepatic section of aqueous group of C. cyminum shows central vein (CV) hepatocytes (HC) and sinusoids (S) with normal sizes, H and E 400 xs

For positive control groups, the present study showed congestion of blood vessels and infiltration of lymphocytes in third and fourth positive control groups (Figures 4 and 5, respectively) as compared with negative control group (Figure 6) that showed normal shape for central vein, hepatocytes and sinusoids.

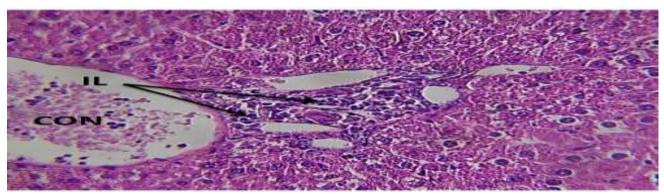


Figure 4: Hepatic section for third group of positive control shows congestion of blood vessels (CON) and infiltration of lymphocytes (IL), H and E 400x

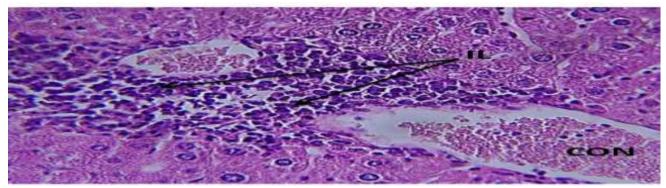


Figure 5: Hepatic section for fourth group of positive control shows congestion of blood vessels (CON) and infiltration of lymphocytes (IL), H and E 400x

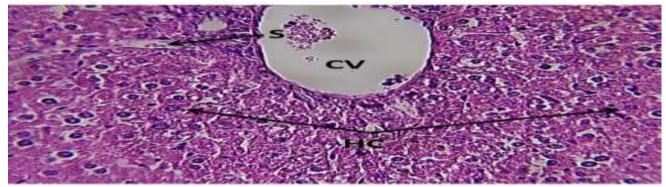


Figure 6: Hepatic section of negative control group shows central vein (CV), hepatocytes (HC) and sinusoids (S) with normal shape, H and E  $400~\mathrm{xs}$ 

The present study agreed with previous studies [44, 45] that detected lymphocytes infiltration and blood vessels congestion with hydatid cyst infection. associated Hossain mentioned et al.. that histopathological effects occur because hydatidosis infection [46]. Polloca et al. Referred to the infiltration in the location of infection, which attributed to parasitic toxins that lead to eosinophils activation [47]. While, other study reported congestion of blood vessels due to weakness in the blood discharge as result to occlusion of hepatic veins by hydatid cyst pressure [48].

#### Spleen

Results in the current study showed normal shape and size for white pulp, red pulp and central artery of all treatment groups (Figures 7, 8 and 9). The main reason for these results is presence of antioxidant compounds in *C. cyminum* extracts, which prevent the binding between hydroxyl free radical with proteins or amino acids [49]. In addition, these active compounds protect DNA of splenic cells and prevent the oxidative stress [50].

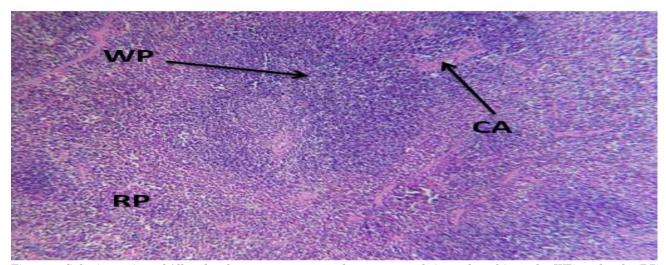


Figure 7: Splenic section of Albendazole treatment group shows a normal tissue for white pulp (WP), red pulp (RP) and central artery (CA), H and E 100x

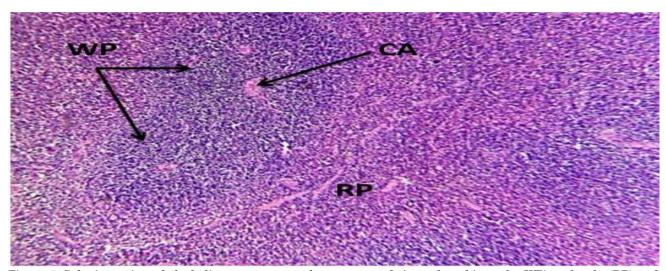


Figure 8: Splenic section of alcoholic extract group shows a normal tissue for white pulp (WP), red pulp (RP) and central artery (CA), H and E 100x

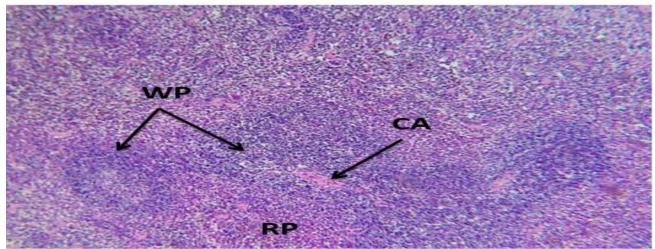


Figure 9: Splenic section of aqueous extract group shows a normal tissue for white pulp (WP), red pulp (RP) and central artery (CA), H and E 100x

About positive control groups, edema and necrotic materials appeared in the third (Figure 10) and fourth (Figure 11) groups as comparison with negative control group that showed normal splenic tissue (Figure 12). These histosplenic changes may attributed to host immune response against hydatid cyst and existence of different cytokines including

interleukin and tumor necrosis factor (TNF), these cytokines lead to blood vessels expanding and somatic fluids accumulation in the peripheral tissue that create edema [51, 52]. While, necrosis can be happens due to the inflammatory reaction between defense cells and outer layer of hydatid cyst [53].

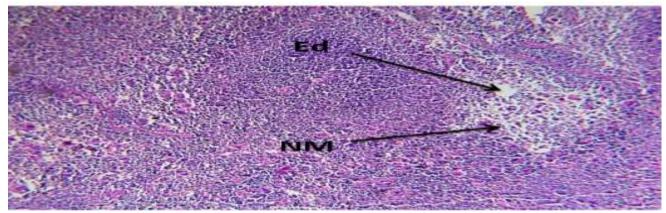


Figure 10: Splenic section of third positive control group shows edema (Ed) and necrotic materials (NM), H and E 100x

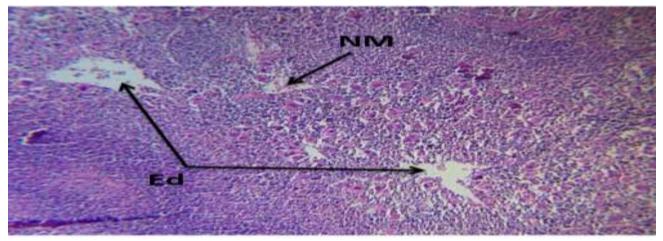


Figure 11: Splenic section of fourth positive control group shows edema (Ed) and necrotic materials (NM), H and E 100x

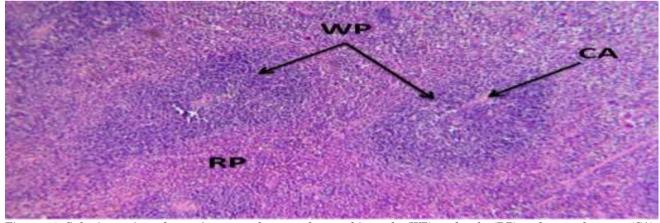


Figure 12: Splenic section of negative control group shows white pulp (WP), red pulp (RP) and central artery (CA) with normal sizes, H and E 100 xs

#### Conclusion

Hydatidosis has pathogenic effects such as rising of total cholesterol and triglyceride, decreasing of heavy density lipoprotein-C, in addition, histological effects for example degeneration, congestion of blood vessels, lymphocytes infiltration, edema and necrosis. These pathogenic effects increase with time progressing. Therefore, *C. cyminum* extracts are useful for attenuating of hydatidosis pathogenic effects because *C. cyminum* has some of active compounds. According to the

present study, *C. cyminum* extracts are better than Albendazole due to no side effects are reported for *C. cyminum* extracts in this study.

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#### References

- 1. Budke CM, Carabin H, Ndimubanzi PC, Nguyen H, Rainwater E, Dickey M, Bhattari R, Zeziulin O, Qian MB (2013) A systematic review worldwide and its associated clinical manifestations. Am. J. Trop. Med. Hyg., 88(6): 1011-1027.
- 2. Eckert TJ, Deplazes P (2004) Biological, epidemiological and clinical aspect of echinococcosis, a zoonosis of increasing concern. Clin. Microbiol. Rev., 17(1): 107-135.
- 3. Siracusano A, Teggi A, Ortona E (2009) Human cystic echinococcosis: old problems and new perspectives. Interdiscip. Perspect. Infect. Dis., 2009: 1-7.
- 4. McManus DP, Zhang W, Li J, Bartley PB (2003) Echinococcosis. The Lancet, 362: 1295-1304.
- 5. Gottstein B (1992) Molecular and immunological diagnosis of echinococcosis. Clin. Microbiol. Rev., 5(3): 248-261.
- 6. Swarna SR, Parija SC (2008) Dot-ELISA for evaluation of hydatid cyst wall, protoscoleces and hydatid cyst fluid antigens in the serodiagnosis of cystic echinococcosis. Rev. Inst. Med. Trop. S. Paulo, 50 (4): 233-236.
- 7. Yasawy MI, Mohamed AE, Al-Karawi MA (1992) Albendazole in hydatid disease: results in 22 patients. Ann. Saudi Med., 12(2): 152-156.
- 8. Devi M, Krishnakumari S (2015) Quantitative estimation of primary and secondary metabolites in hot aqueous extract of *Pleurotus sajor caju*. J. Pharmacog. Phytochem., 4(3): 198-202.
- 9. Ashayerizadeh O, Dastar B, Shargh MS (2009) Use of garlic (*Allium sativum*), black cumin seeds (*Nigella sativa L.*) and wild mint (*Mentha longifolia*) in Broilert chickens diets. J. Anim. Vet. Adv., 8 (9): 1860-1863.
- 10. Smyth JD (1985) In vitro culture of *Echinococcus* spp. Proc. 13th edn., In. Corg. Hydit. Madrid, 84-95.
- 11. Rotunno CA, Kammerer WS, Esandi M, Cerejido M (1974) Studies on the permeability to water, sodium and chloride of the hydatid cyst of *E. granulosus*. J. Parasitol., 604: 613- 620.

- 12. Smyth JD, Barrett NJ (1980) Procedure for testing the viability of human hydatid cysts following surgical removal, especially after chemotherapy. Trans. Roy. Soci. Trop. Med. Hyg., 74(5): 649-652.
- 13. Campbell DH, Garvey JS, Cremer NE, Sussdorf DH (1964) Methods in immunology, 2nd edn., Benjamin Inc., New York, USA, 263.
- 14. Theodorides Y, Frydas S, Rallis T, Adamama-Moraitou K, Papazahariadou R, Batzios C, Conti P (2001) MCP-1 and MIP-2 levels during *Echinococcus granulosus* infections in mice. J. Helminthol., 75: 205-208.
- 15. Myers RL (1995) Immunology, a laboratory manual, 2<sup>nd</sup> ed., W.C. Brown Communications, Dubuque, 133.
- 16. Anadol D, Özcelik U, Kiper N, Gocmen A (2001) Treatment of hydatid disease. Paediatr. Drugs., 32: 123-135.
- 17. Jinesh VK, Jaishree V, Badami SH, Shyam W (2010) Comparative evaluation of antioxidant properties of edible and non-edible leaves of *Anethum graveolens* Linn. Indian J. Natur. Prod. Resour., 1(2): 168-173.
- 18. Ahmed I, Mehmood Z, Mohammad F (1998) Screening of some Indian medicinal plants for their antimicrobial properties. J. Enthnopharm., 62: 183-193.
- 19. Harborne JB (1973) Phytochemical methods: a guide to modern techniques of plant analysis. Chapman and Hall Ltd., London, 278.
- 20. Mahmoud MJ (2008) Chemistry of medicinal plants. Anwar Dijla, Bagdad, Iraq, 13-16 pp.
- 21. Kokate CK, Purohit AP, Gokhale SB (2009) Pharmacognosy. 17<sup>th</sup> edn. Nirali Prakashan, 231.
- 22. Rashant T, Bimlesh K, Mandeep K, Gurpreet K, Harleen K (2011) Phytochemical screening and Extraction. Int. Pharm. Sci., 11: 98-106.
- 23. Harborne JB (1984) Phytochemical methods. 2<sup>nd</sup> edn., Chapman and Hall, New York, USA., 300.
- 24. Bancroft J, Stevens A (1987) Theory and practice of histological technique. *Edinburgh and London*, 482-502.

- 25. Landau S, Everitt BS (2004) A handbook of statistical analyses using SPSS. Chapman and Hall/CRC., 329.
- 26. Hussein HKA, Kadhem NH, Abbod ZH (2007) Study of the biological activity of aqueous extract of *Cuminum cyminum* L. and *Hibiscus sabdariffa* L. and detection of some active groups in them. J. Kerbala Univ., 5(1): 65-72.
- 27. Ageena SJ, Hindi MG, Yahya AA (2009) Effect of some ethanolic plant extracts on the inhibition of some types of pathogenic bacteria and causing spoilage of food. Iraqi J. Market Res. Consumer Prot., 1(2): 1-14.
- 28. Al-Myah ARAA, Al-Mansour N, Al-Dhahir AHS (2011) Effect of some plants extracts on the mortality of the larval mosquitoes Culex pipiens molestus Forskal. Basrah J. Sci., 29(1): 47-61.
- 29. Kolodziej H, Kiderlen AF (2005) Antileishmanial activity and immune modulatory effects of tannins and related compounds on *Leishmania* parasitised RAW 264.7 cells. Phytochemistry, 66(17): 2056-2071.
- 30. Muthamma MKS, Dholakia H, Kaul TP, Vishveshwaraiah P (2008) Enhancement of digestive enzymatic activity by cumin (*Cuminum cyminum* L.) and role of spent cumin as a bionutrient. Food Chem., 110: 678-683.
- 31. Dua A, Gaurav G, Balkar S, Mahajan R (2013) Antimicrobial properties of methanolic extract of Cumin (*Cuminum cyminum*) seeds. Int. J. Res. Ayurveda Pharm., 4(1): 104-107.
- 32. Al-Quraishi MA, Shaalan NN, Almusawi HS (2015) Study the effect of Artemisia Herba-alba extracts in adult and larval stages of *Echinococcus granulosus* parasite *in vivo* and *in vitro*. Int. J. Curr. Microbiol. App. Sci., 4(8): 267-282.
- 33. Hayden MR, Tyagi SC (2005) Isolated low high density lipoprotein cholesterol (HDL-C): implications of global risk reduction. Case Rep. Sys. Sci. Rev., 4: 4.
- 34. Jemai H, Fki I, Bouaziz M, Bouallagui Z, El-Feki A, Isoda H, Sayadi S (2008) Lipid lowering and antioxidant effects of hydroxytyrosol and its triacetylated derivative recovered from Olive tree leaves in cholesterol-fed rats. J. Agric. Food Chem., 56(8): 2630-2636.

- 35. Feryal S, Hasan F Ali S (2011) Effect of alcoholic *Anastatica hirochuntica* extract on some biochemical and histological parameters in alloxan induced diabetic rats. Iraqi J. Sci., 52(4): 445- 455.
- 36. Martin DW, Mayes PA, Rodwell VW, Cranner DK (1995) Biochemistry 20<sup>th</sup> ed., Lange Medical publications, Los Altos, California, USA., 299.
- 37. Tan KCB, Shin SWM, Chu BYM (2000) Effects of gender, hepatic lipase gene polymorph and type 2 diabetes mellitus on hepatic lipase activity in Chinese. Athersclerosis, 157(1): 133.
- 38. Njoroge E, Mbihi P, Wachira T, Gathuma J, Gathura P, Maitho TE, Magambo J, Zeyhle E (2005) Comparative study of Albendazole and Oxfendazole in the treatment of cystic echinococcosis in sheep and goats. Int. J. Appl. Res. Vet. Med., 3(2): 100.
- 39. Al-Meyah SH, Al-Hilfy AA, Abu-Mejdad NM (2010) Evaluation of treatment effects of vaccine and Albendazole against *Echinococcus granulosus* L. *in vitro* and *in vivo*. J. Basrah Res. Sci., 36(15B): 4.
- 40. Rawat AKS, Methrotra S, Tripothi SC, Shome U (1997) Hepatoprotective activity of Boerhavia diffuse L-roots a popular Indian ethno medicine. J. Ethno., 56:61-66.
- 41. Oriskwe OE, Afonnc OS, Chude MA, Ob E, Dioka CE (2003) Sub chronic toxicity studies of the aqueous extract of *Bacrhara diffusa* leaves. Hlth. Sci., 49(6): 441-447.
- RP, 42. Hewawasam Jagatilaka KAPW, Pathirara C, Mudduwa LKB (2004)Hepataprotective effect of **Ephtes** divaricate extract on carbon tetrachloride induced hepatatxicry in mice. Indian Sci. Med. Res., 120: 30-34.
- 43. Ali N, Rashid S, Nafees S, Hasan K, Sultana S (2014) Beneficial effects of Chrysin against Methotrexate-induced hepatotoxicity via attenuation of oxidative stress and apoptosis. Mol. Cell Biochem., 385: 215-223.
- 44. Das DK, Bhambhani S, Pant CS (1995) Ultrasound guided fine needle aspiration, cytology diagnosis of hydatid disease of the abdomen and thorax. Diag. Cytopathol., 12: 173-176.

- 45. Al-Shanawi FA, Baker NN (2011) Study the effect of the mixture alcoholic extract of *Peganum harmala* seeds and cones of *Cupressus sempervirens* and their effect on viability of protoscolices of *Echinococcus granulosus in vivo*. J. Biotechnol. Res. Cen., 5(2): 44-52.
- 46. Hossain A, Shanta IS (2007) Pathological investigation of liver of the slaughtered buffaloes in Barisal district. Bangl. J. Vet. Med., 5: 81-85.
- 47. Pollaco S, Nicholas WL, Mitche GF, Stewart AC (1978) T-cell dependent collagenous encapsulating response in the mouse liver to Mesocestodoides corti. Int. J. Parasitol., 8:457-467.
- 48. Mir SH, Abdul-Baqui BRC, Darzi MM, Abdul-Wahid S (2008) Biochemical and histomorphological study of Streptozotocin-Induced Diabetes Mellitus in Rabbits. Pakistan J. Nut. 7(2): 359-364.
- 49. Cadenas E, Davies KJ (2000) Mitochondrial free radical generation, oxidative stress and free aging. Radic. Biol. Med., 29(3-4): 222-230.

- 50. Abd Elhalim SA, Sharada HM, Abulyazid I, Aboulthana WM, Abd Elhalim ST (2017) Ameliorative effect of carob pods extract (*Ceratonia siliqua* L.) against Cyclophosphamide induced alterations in bone marrow and spleen of Rats. J. Appl. Pharm. Sci., 7(10): 168-181.
- 51. Rautenschlein S, Yeh HN, Sharma JM (2002) The role of T cells in protections by an inactivated infectious bursal disease virus vaccine. Vet. Immunol. Immunopathol., 89(3-4): 159-167.
- 52. Tizard IR (2004) Macrophages and the later stages of inflammation. In: Veterinary immunology an introduction. 7th ed. Saunders, 401.
- 53. Al-Fartosi KG, Al-Abady FA, Al-Badry FA (2014) Effect of camel's milk on some histological changes of testis and spleen of male rats treated with cow's livers infected by *Fasciola gigantica*. J. Thi-Qar Univ., 2(4): 61-69.

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